

# Designing A Better Assay- HLA-DR Binding Assays as a Tool for Immunogenicity Screening: Developing the Best Practices for Determining Immunogenicity

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## Introduction

### Purpose

- As generic drugs and protein therapeutics become more readily available, anti-drug immunogenicity becomes a greater concern.
- HLA-type has been correlated with immunogenicity and several in silico and in vitro methods exist for screening protein therapeutics for the presence of HLA-binding peptides. (Cousens et al 2006)
- While multiple in vitro methods for assessing HLA binding are in use, **not all are created equally**.
- Knowing which methods provide the most accurate and reliable results is important for assessing the assay's reliability.

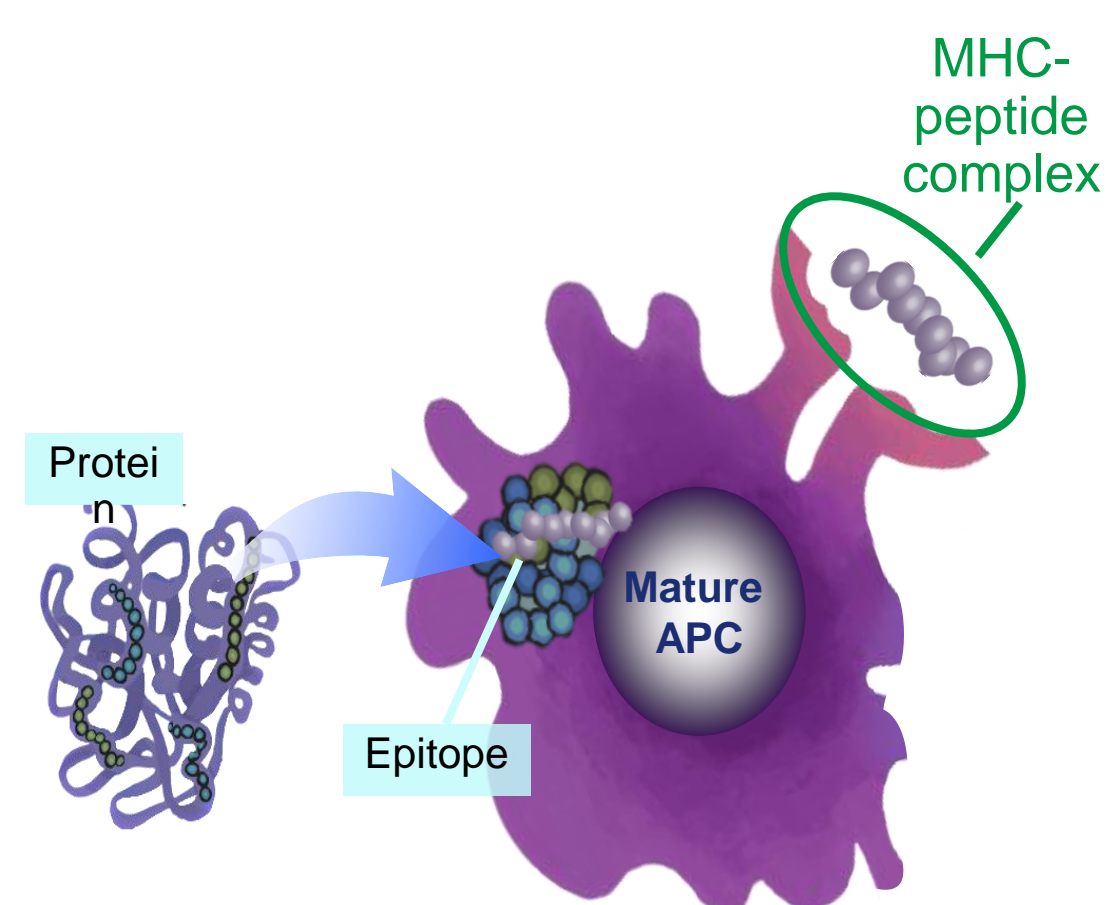
### Objectives

- To compare and contrast several methods in use to measure HLA-binding in vitro
  - Single concentration vs dilution curve
  - Overlapping peptide libraries
    - The importance of properly centered peptides

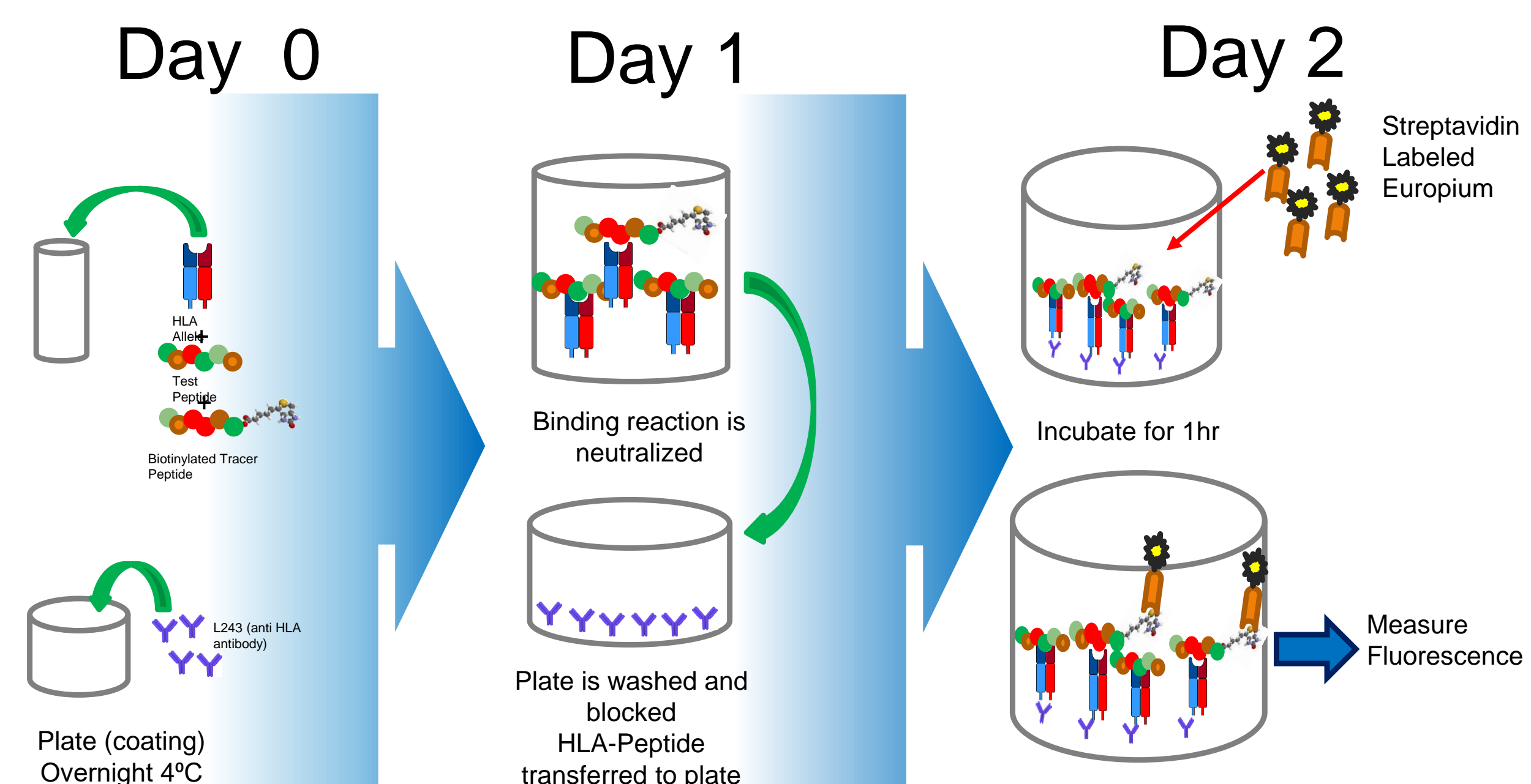
### Methods

#### Immunogenicity assessment:

- EpiMatrix** takes a protein sequence and parses it into overlapping 9-mer frames. Peptides are analyzed for their ability to bind 8 MHC-II supertype alleles.



- HLA Binding Assay:** Peptides that are predicted to bind MHC are synthesized and assayed over a range of 7 concentrations in our HLA-binding assay. In brief, peptides are incubated overnight with soluble HLA and a biotin labeled competitor of moderate affinity. On day 2, the reaction is halted and the mixture is transferred to a plate coated with a pan anti-HLA antibody. On day 3, plates are developed by the addition of streptavidin-Europium and fluorescence is measured.



## Assay Sensitivity: Single Concentration vs Dose Range

Figure 1a

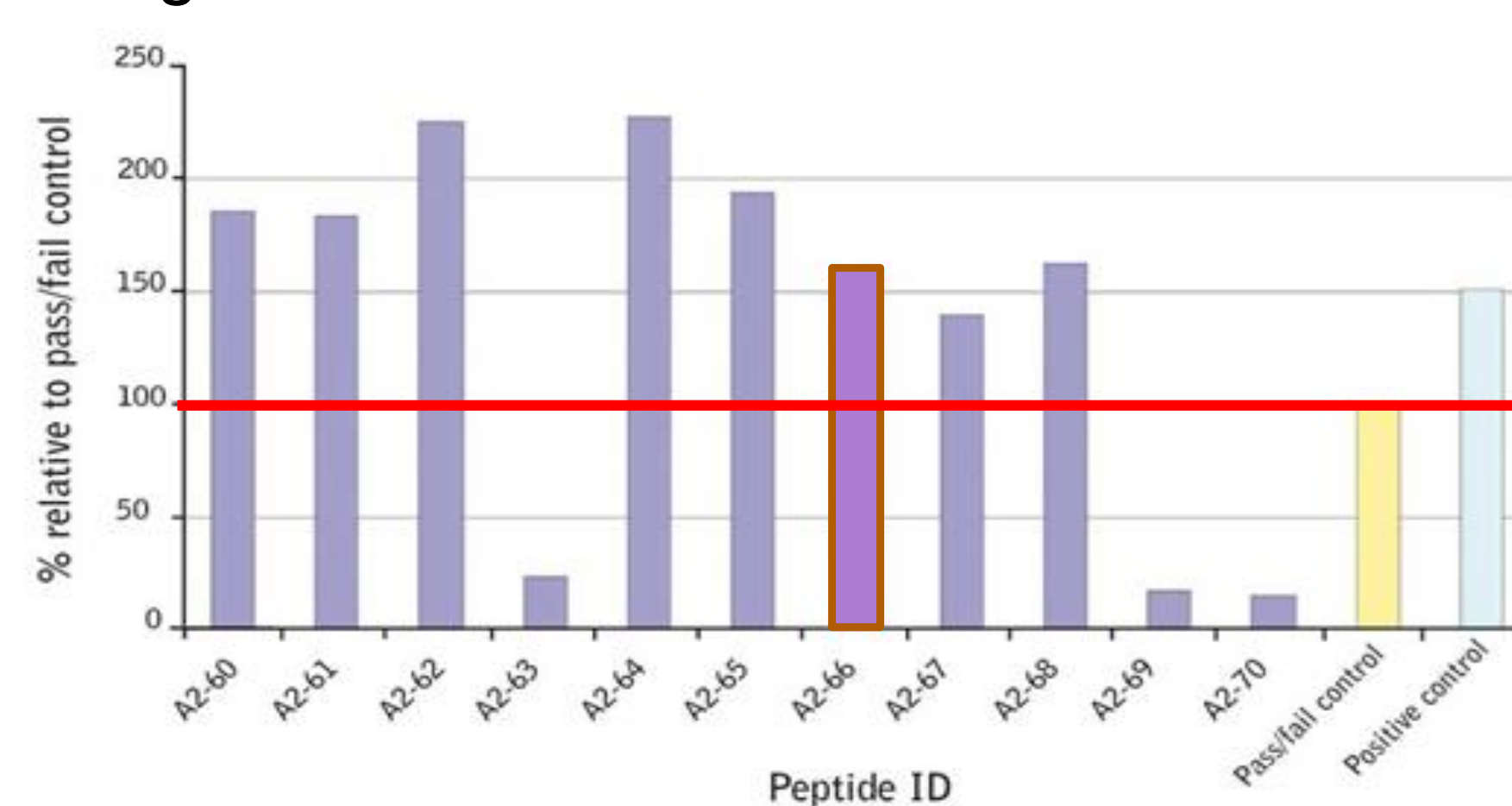
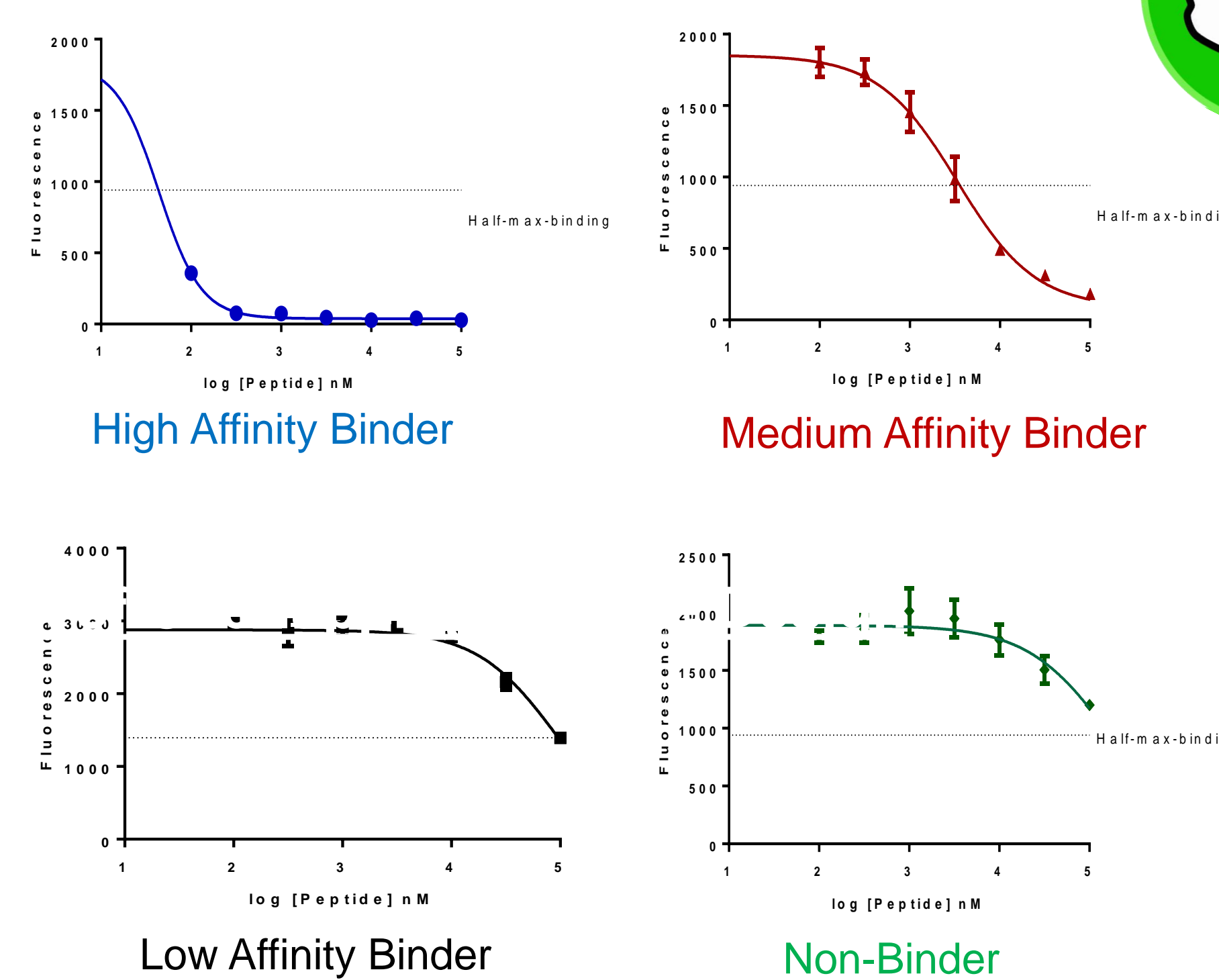


Figure 1b



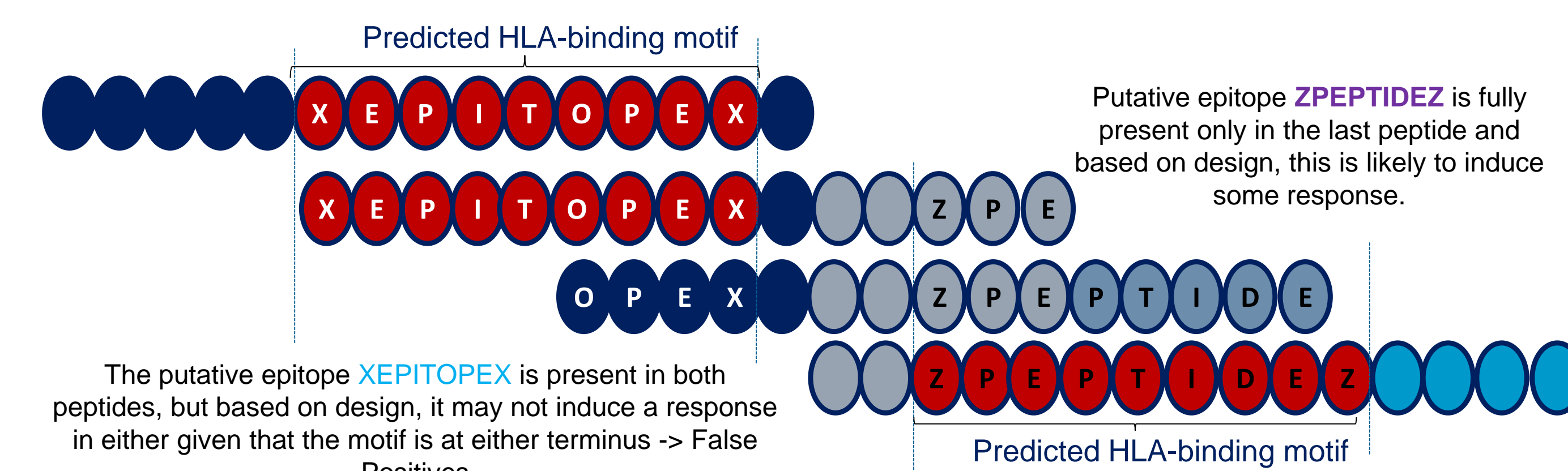
Poor Design

Best Design

Non-binder (No dose-dependent inhibition)
Negligible Affinity (100,000 nM < IC <sub>50</sub> )
Low Affinity (10,000 nM < IC <sub>50</sub> < 100,000 nM)
Moderate Affinity (1,000 nM < IC <sub>50</sub> < 10,000 nM)
High Affinity (100 nM < IC <sub>50</sub> < 1,000 nM)
Very High Affinity (IC <sub>50</sub> < 100nM)

Figure 1. (a) Assaying peptides at a single concentration requires measuring binding relative to a pass/fail control. Peptides that don't meet this cutoff value (red line) would be considered "non-binding" and excluded from further screening why they could contribute to the immunogenicity of the drug. (b) Measuring peptide binding over a range of concentrations provides information about the peptide-HLA affinity and allows us to classify peptides as either strong, moderate, weak or non-binders.

## The Drawback of Overlapping Peptide Design

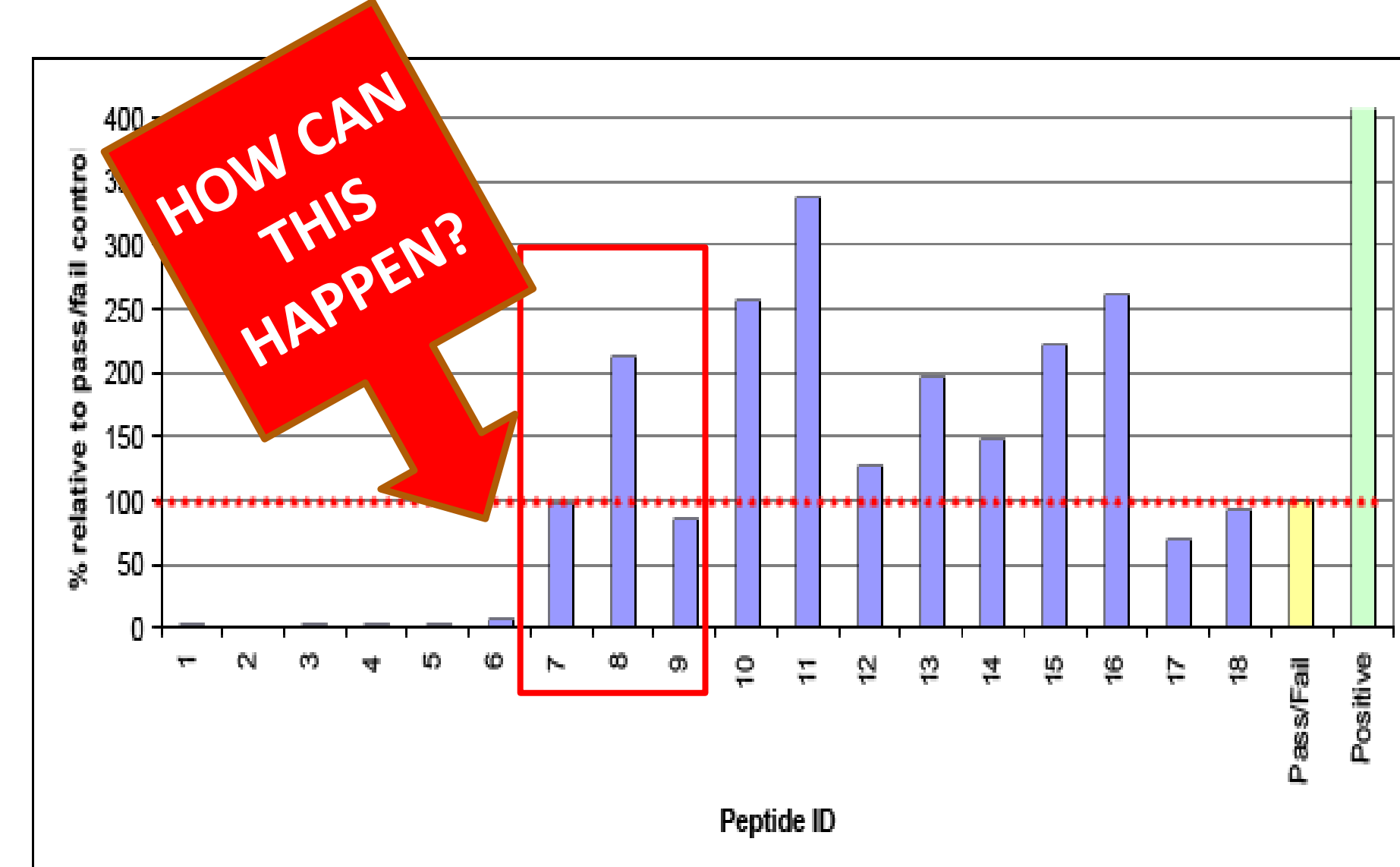


The putative epitope XEPITOPEX is present in both peptides, but based on design, it may not induce a response in either given that the motif is at either terminus -> False Positives

Putative epitope ZPEPTIDEZ is fully present only in the last peptide and based on design, this is likely to induce some response.

The truncated epitope (ZPEPTIDE) though not predicted to bind, could also induce a response if residue at position 1 ("Z") is a strong P1 binding anchor, -> False Negative

Overlapping (OL) peptide libraries contain binding motifs that are not properly centered. Peptides that bind to HLA Class II molecules are stabilized by flanking residues. When considering synthetic peptides for in vitro testing and validation, **poorly centered HLA-binding motifs may result in absence of binding or T cell response** despite the presence of an HLA binding motif. Synthesizing OL peptide libraries is also costly and time consuming.



An example of overlapping peptide data is shown below. In this example, each peptide overlaps its neighbor by 14 amino acids therefore peptides 7 and 9 should contain the same nine-mer found in peptide 8 that bind to HLA.

Poor Design

Best Design

ORIGINAL	OPTIMIZED	ORIGINAL	OPTIMIZED
EpiMatrix Cluster Detail Report RH36-50 Cluster: 36	EpiMatrix Cluster Detail Report RH36-50MOD Cluster: 33	EpiMatrix Cluster Detail Report 07_IL46-60 Cluster: 46	EpiMatrix Cluster Detail Report 08_IL46-60MOD Cluster: 43
Summarized Results: 36 binders	Summarized Results: 33 binders	Summarized Results: 46 binders	Summarized Results: 43 binders
Strong binding motif (EpiBar) located at flanks	Optimized Peptide has a centered binding motif	Four more binders observed with repeat assay	With optimized version, we find more binders with stronger affinities
IC50 values obtained qualify as binders according to EpiVax criteria	With optimized version, we find binders with stronger affinities		
Publication Results (HLA-DR alleles: DR1, DR4, DR7, DR11, DR15)	Optimized Peptides Results (EpiVax) (HLA-DR alleles: DR1, DR4, DR7, DR11, DR15)		
Positive Predictive Value (excluding RL46-60): 22%	Positive Predictive Value (including RL46-60): 83%		



In a study by Hamze et al ("original") we observed a poor correlation between in silico predictions and HLA-binding. When peptides were properly centered ("optimized"), we see improved correlation between the in silico and in vitro results. **Optimizing peptide design leads to stronger correlation and higher binding affinities**

Note: Only a subset of the data is shown here and we are only reporting data for optimized peptides

## Conclusions

- Assay design is critical for correlating in silico and in vitro HLA binding data.
- To draw the most accurate conclusions on immunogenicity towards biologics, the industry must adopt a consistent and standard set of best practices for peptide and assay design.

## References

Cousens LP, Najafian N, Mingozi F, et al. In Vitro and In Vivo Studies of IgG-derived Treg Epitopes (Tregitopes): A Promising New Tool for Tolerance Induction and Treatment of Autoimmunity. *J Clin Immunol* 2013; 33: 43-49.

Steere AC, Kitz W, Drouin EE, et al. Antibiotic-refractory Lyme arthritis is associated with HLA-DR molecules that bind a Borrelia burgdorferi peptide. *J Exp Med* 2006; 203: 961-971.

Hamze M, Meunier S, Karle A, et al. Characterization of CD4 T Cell Epitopes of Infliximab and Rituximab Identified from Healthy Donors. *Frontiers in Immunology*. 2017;8:500.

