

From infectious disease to personalized cancer vaccines:

Regulatory T cell epitopes determine patient outcomes and suppress neoantigen responses CHICAGO



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Abstract

Precision cancer immunotherapy has proven to effectively control the tumor of patients in multiple clinical trials. However, the selection of immunogenic T cell neo-epitopes using traditional methodologies remains a challenging exercise. Poor vaccine performance may be due to inclusion of mutated epitopes cross-conserved with self-epitopes recognized by regulatory (Treg), anergic, or deleted T cells. In addition, most cancer vaccine studies focus on the selection of CD8 T cell neo-epitopes while overlooking CD4 T cell neoepitopes.

We have developed Ancer, an integrated and streamlined neo-epitope selection pipeline, that accelerates the selection of both CD4 and CD8 T cell neo-epitopes. Ancer leverages EpiMatrix and JanusMatrix, predictive algorithms that have been extensively validated in prospective vaccine studies for infectious diseases [1]. Distinctive features of Ancer are its ability to accurately predict Class II HLA ligands and to identify tolerated or Treg epitopes.

Ancer was evaluated on data from the BLCA bladder cancer cohort hosted at The Cancer Genome Atlas (TCGA) database. An initial analysis was carried out on a representative set of eleven patients to derive the best vaccine candidate sequences from high-quality missense mutations. A median number of 24 [interquartile range: 15-64] candidate sequences were generated for each patient under study. This initial analysis demonstrates the capacity of Ancer to define a sufficient number of candidate sequences for vaccinating bladder cancer patients in a precision immunotherapy setting.

We also assessed Ancer's ability to explain patients' disease free survival (DFS) times. While BLCA patients with DFS greater and lower than 3 months could not be distinguished by their tumor mutational burden (p=0.19), nor by their load of missense mutations (p=0.15), the number of mutations identified as highly distinct from self, based on Ancer, significantly segregated this two patient populations (p=0.02). Similar results were obtained when studying BLCA patients with DFS greater and lower than 6 months. These observations suggest that defining the number of true non-self mutations using Ancer may represent a novel biomarker for more robust anti-tumor immune response and higher likelihood of disease-free survival.

Our preliminary analysis of the BLCA cohort from the TCGA database showcases the value of Ancer in clinical settings. Our next step will be to investigate whether Ancer-defined neoepitope load will serve as a biomarker for prognosis and response to therapy in the full BLCA cohort.

Mutanome-Directed Cancer Immunotherapy Based on 20 Years of Experience in Epitope Mapping Neo-Antigen Discovery Ancer™ Ancer[™] Platform: the "Answer" to Cancer Triage mutations based on NGS metadata and transcriptomics Scan for HLA Matched CD4 and CD8 Epitopes with EpiMatrix® Screen for "Non-Self" Neo-Epitopes with JanusMatrix™ Rank and Select Peptide Candidates Identification and removal of Treg epitopes using JanusMatrix™ JanusMatrix™ identifies Treg epitopes can reduce Antigens optimized with IFN_γ ELISpot responses cross-reactive and JanusMatrix are more regulatory T cell (Treg) observed for peptide pools [3]. immunogenic [4]. epitopes. 5-fold increase

Peptide pool

Peptide pool

JanusMatrix-derived

H7: wild-type H7 antigen

Opt1: Optimized H7 antigen w/o JanusMatrix-derived Treg epitope

Accurate and identification of CD4 and CD8 epitopes using EpiMatrix®

CD4 T cell epitopes

The predictive accuracy of EpiMatrix® is routinely evaluated by testing predicted HLA ligands in in vitro HLA binding assays.

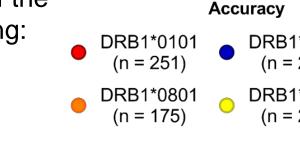
EpiMatrix Class II HLA predictions are 74% accurate when tested in in vitro HLA binding assays, with an average observed PPV of

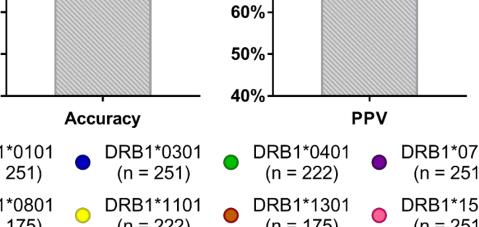
EpiMatrix® CD4 predictions and its associated tools are routinely used and trusted by 9 of the top 10 pharmaceutical companies, including:











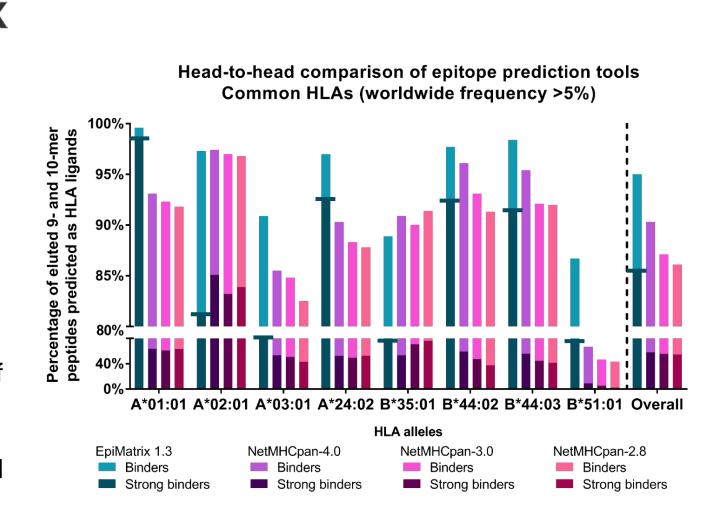
Accuracy and PPV of EpiMatrix Class II predictions

Prospective selection of HLA ligands

CD8 T cell epitopes

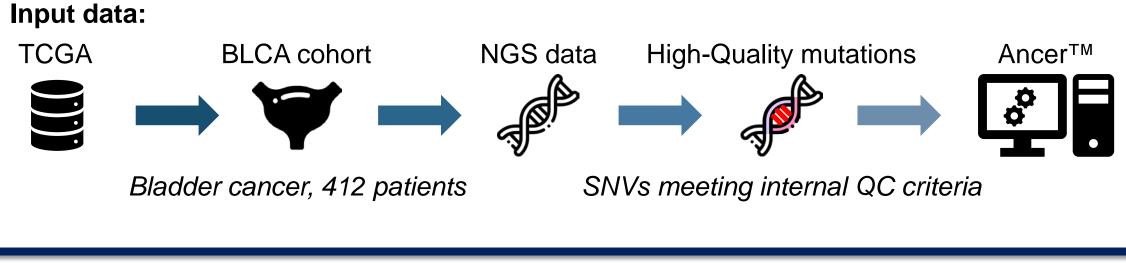
Analysis of eluted peptide dataset [2]: 95% of eluted 9- and 10-mers were predicted to bind to HLA according to EpiMatrix®, while only ~88% of ligands were accurately recalled by NetMHCpan.

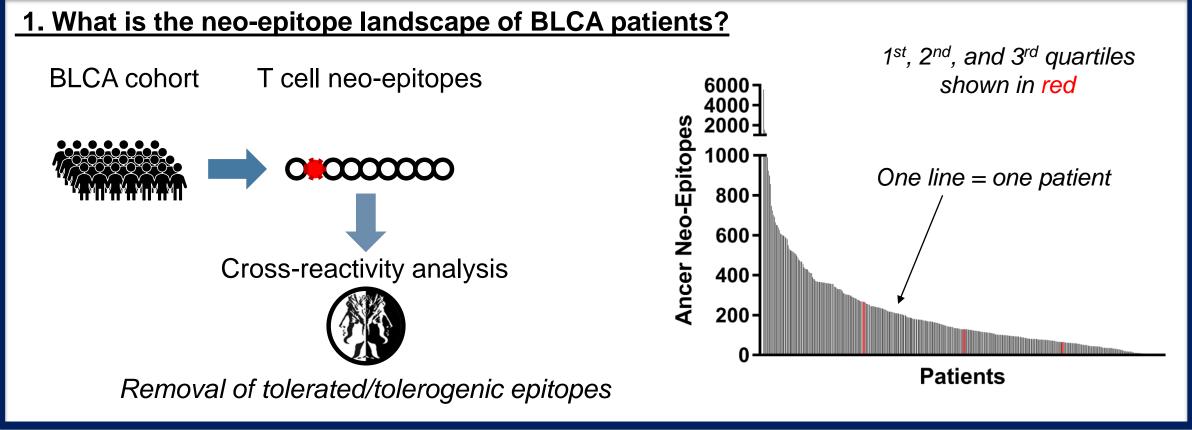
The majority of eluted peptides (85% of the dataset) are strong EpiMatrix® binders, while less than 56% of all peptides are high affinity binders based on NetMHCpan predictions.

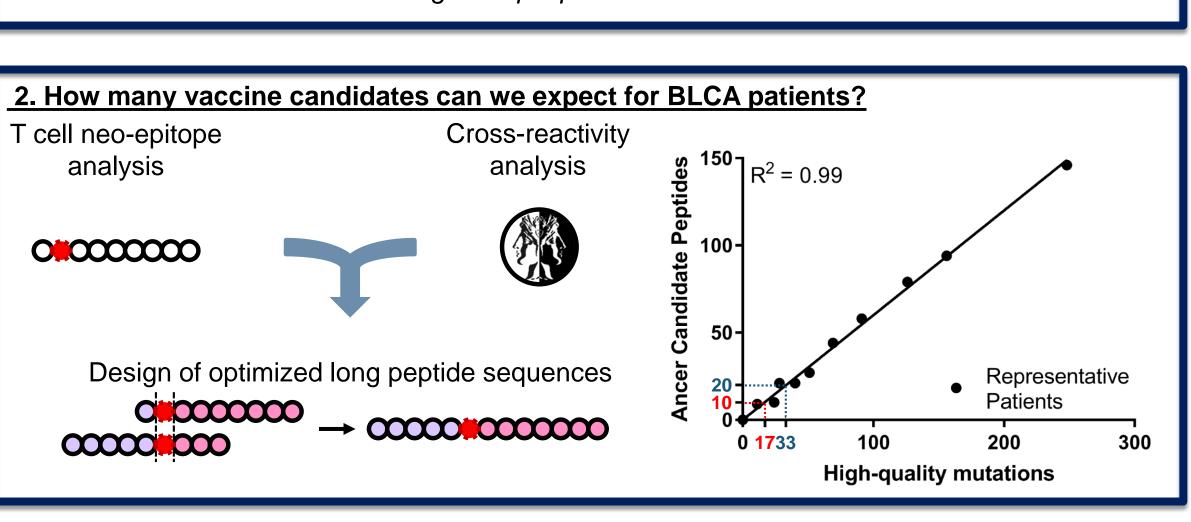


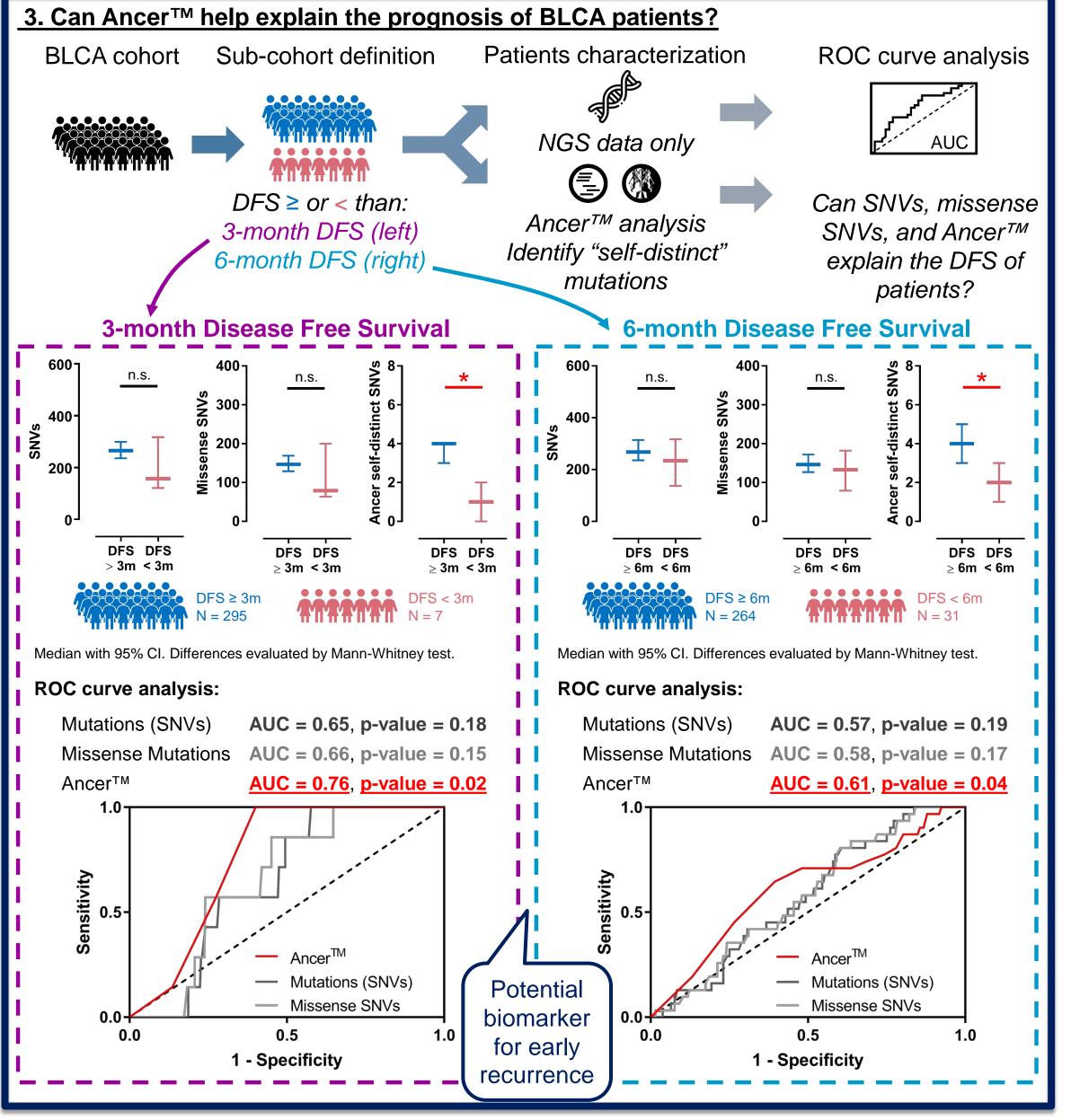
Ancer™ Retrospective Study of the TCGA Database: Bladder Cancer (BLCA) cohort

Ancer™ was used to analyze bladder cancer NGS data from the TCGA database and to answer the following questions.



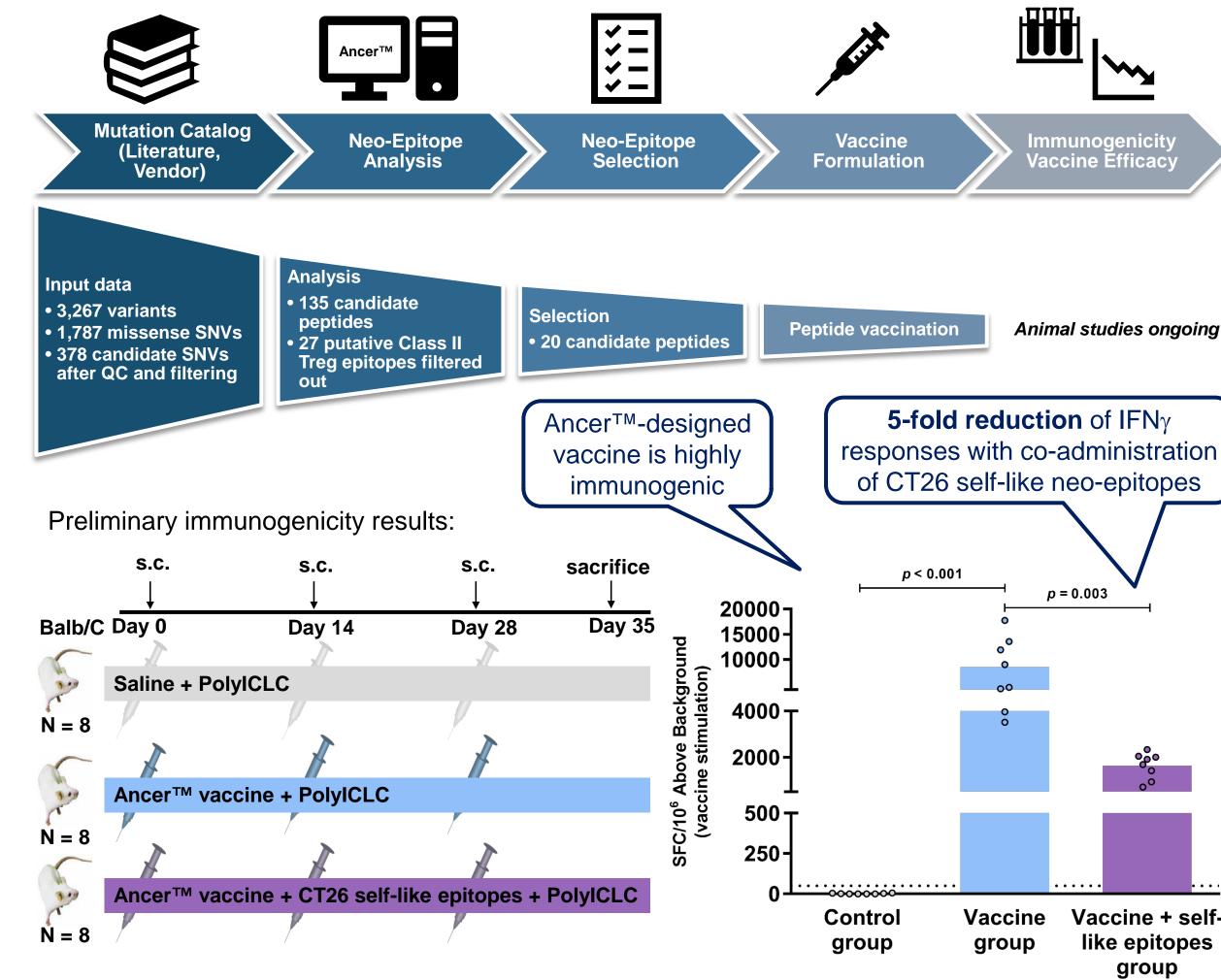






Ancer™ Prospective CT26 Studies

Ancer™ is central to the design of prospective studies using the CT26 syngeneic mouse models.



Splenocytes were collected at day 35 and restimulated in IFN_γ ELISpot assay with the 20 vaccine peptides.

References and Acknowledgments

Strong IFNy responses are observed in the vaccinated group after restimulation compared to the control group. Co-administration of Ancer™-derived CT26 self-like neo-epitopes identified with JanusMatrix™ significantly reduce IFN_γ responses by 5-fold.

Conclusions

- Analysis of the MHC- and TCR-facing residues of T cell epitopes by JanusMatrix™ enables prediction of epitope phenotype.
- EpiVax's immunogenicity screening tools (EpiMatrix® and JanusMatrix™) are integrated into the Ancer™ platform for streamlined designs of personalized cancer vaccines.
- While only HLA Class I alleles were available for TCGA bladder cancer patients, results reveal that Ancer™ may predict likelihood of disease-free survival. Follow-up analyses are planned where HLA Class II will be considered.
- Ancer[™]-derived vaccines are highly immunogenic in a CT26 mouse model. Our results showcase that inclusion of Treg neo-epitopes in cancer vaccines can downregulate immune responses.
- 1) Moise L. et al., Hum Vaccin Immunother. 2015;11(9):2312-21.
- 2) Abelin J. et al., Immunity. 2017 46, 315–326 3) Wada Y. et al., Sci Rep. 2017 Apr 28;7(1):1283
- 4) Liu R. et al., Hum Vaccin Immunother. 2015 11:9 2241-2252

The results shown here are in part based upon data generated by the TCGA Research Network: http://cancergenome.nih.gov/.

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