

# Assessing the risk of alloimmune responses by scanning the human proteome

Adrian Shepherd

ISMB & Biological Sciences, Birkbeck, University of London

Immunogenicity and Tolerance Seminar, Amsterdam

15<sup>th</sup> November 2019



# Overview

- Birkbeck and my research group
- Missense mutation haemophilia A
- Proteome scanning: our strategy for identifying when self tolerance is likely to be broken
- Other applications, notably to organ transplantation

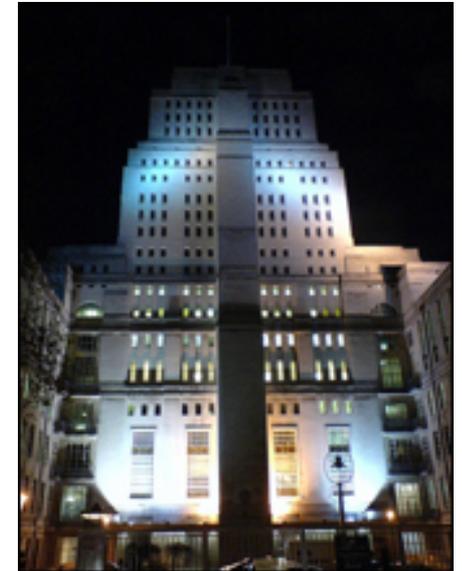
# About Birkbeck

Founded in **1823**

University of London's only specialist provider of **part-time** higher education

Nobel Laureates **Aaron Klug** and **Derek Barton** worked in the School of Crystallography

Biological Sciences is part of a joint research institute with UCL: **ISMB**



*Motto: In nocte consilium*



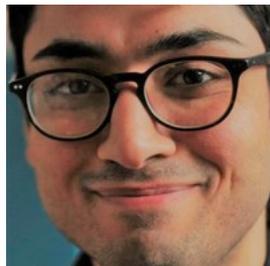
# Research overview

## Immunoinformatics:

- Antibodies (repertoire analytics, MD simulation, deep learning)
- B-cell epitopes (predicting antigenic escape)
- T-cell epitopes (allo-immunity, heterologous immunity)

## Antigens of interest:

- Protein therapeutics (e.g. tFVIII)
- Viruses (e.g. HCV, HIV, IAV)
- Bacteria (e.g. B. anthracis)
- Cancer (HCC)



# The AIRR Community

Established in 2015 at first of 4 Community Meetings:

- Devising relevant **standards** + **guidelines** about good practice
- Obtaining, analyzing, curating and comparing/sharing NGS **AIRR datasets**
- **Validating tools** for AIRR data analysis
- **Legal and ethical issues** involving the use and sharing of AIRR data sets derived from humans

A chapter of the **Antibody Society** since 2018.



# AIRR Community recommendations



## AIRR Community Standardized Representations for Annotated Immune Repertoires

OPEN ACCESS

Jason Anthony Vander Heiden<sup>1†</sup>, Susanna Marquez<sup>2</sup>, Nishanth Marthandan<sup>3</sup>, Syed Ahmad Chan Bukhari<sup>2</sup>, Christian E. Busse<sup>4</sup>, Brian Corrie<sup>5</sup>, Uri Hershberg<sup>6,7,8</sup>, Steven H. Kleinstei<sup>2,9</sup>, Frederick A. Matsen IV<sup>10</sup>, Duncan K. Ralph<sup>10</sup>, Aaron M. Rosenfeld<sup>6</sup>, Chaim A. Schramm<sup>11</sup>, The AIRR Community<sup>‡</sup>, Scott Christley<sup>12\*†</sup> and Uri Laserson<sup>13\*</sup>

*Vander Heiden et al., Frontiers in Immunology, 2018*

Adaptive Immune Receptor Repertoire Community recommendations for sharing immune-repertoire sequencing data

*Rubelt et al., Nature Immunology, 2017*

Florian Rubelt<sup>1,21</sup>, Christian E Busse<sup>2,21</sup>, Syed Ahmad Chan Bukhari<sup>3,21</sup>, Jean-Philippe Bürckert<sup>4</sup>, Encarnita Mariotti-Ferrandiz<sup>5</sup>, Lindsay G Cowell<sup>6</sup>, Corey T Watson<sup>7</sup>, Nishanth Marthandan<sup>8</sup>, William J Faison<sup>9</sup>, Uri Hershberg<sup>10</sup>, Uri Laserson<sup>11</sup>, Brian D Corrie<sup>12,13</sup>, Mark M Davis<sup>1,14</sup>, Bjoern Peters<sup>15</sup>, Marie-Paule Lefranc<sup>16</sup>, Jamie K Scott<sup>8,12,17</sup>, Felix Breden<sup>12,13</sup>, The AIRR Community<sup>18</sup>, Eline T Luning Prak<sup>19,22</sup> & Steven H Kleinstei<sup>3,20,22</sup>

High-throughput sequencing of B and T cell receptors is routinely being applied in studies of adaptive immunity. The Adaptive Immune Receptor Repertoire (AIRR) Community was formed in 2015 to address issues in AIRR sequencing studies, including the development of reporting standards for the sharing of data sets.

# AIRR Community resources

**OGRDB** <https://ogrdb.airr-community.org/>  
Lees et al., *NAR*, 2019

**VDJbase** <https://www.vdjbase.org/>  
Omer et al., *NAR*, 2019

**sumrep** <https://github.com/matsengrp/sumrep>  
Olson et al., *Frontiers in Immunology*, 2019

**OGRDB**

**VDJbase**



# Germline reference set errors

Immunology & Cell Biology

 Australian and New Zealand  
SOCIETY FOR IMMUNOLOGY INC.

Outstanding Observation

**Many human immunoglobulin heavy-chain IGHV gene polymorphisms have been reported in error**

Yan Wang, Katherine J L Jackson, William A Sewell, Andrew M Collins 

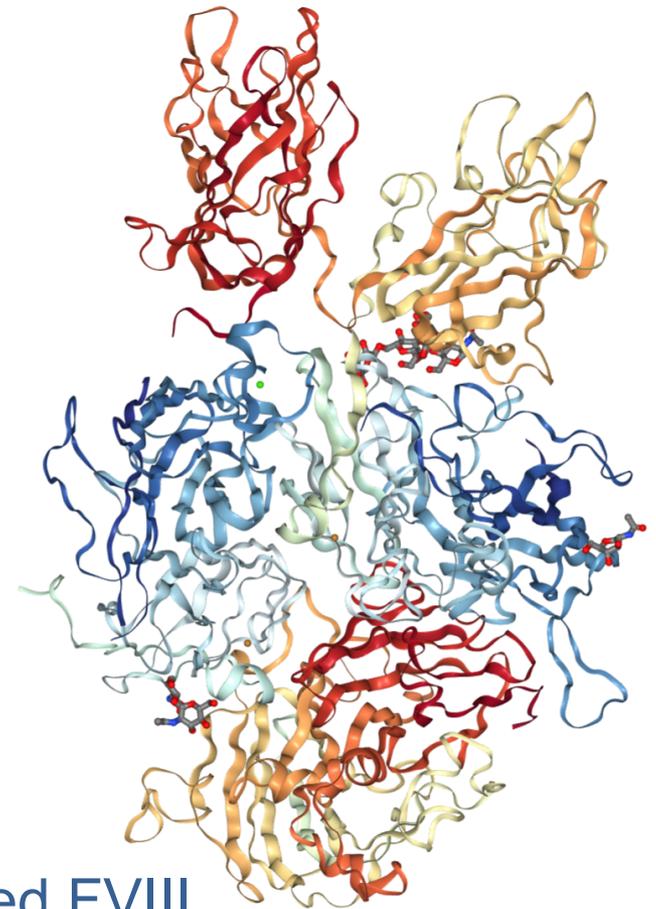
First published: 27 November 2007 | <https://doi.org/10.1038/sj.icb.7100144> | Citations: 30

Of the **226 human IGHV genes** in the **IMGT** database in 2007, **at least 104 were likely to contain sequencing errors**. Almost all of these genes are still there.

# Haemophilia and Inhibitors

# Haemophilia A

- X-linked disease affecting **1 in 5000 men**
- Caused by defect in clotting **factor VIII** (FVIII), part of the coagulation cascade
- Standard treatment is **replacement FVIII** (very expensive recombinant therapeutic)



B-domain-deleted FVIII,  
PDB 3CDZ

# Haemophilia A gene therapy trial

## NEWS

[Home](#) | [UK](#) | [World](#) | [Business](#) | [Politics](#) | [Tech](#) | [Science](#) | [Health](#) | [Family & Education](#)

### Health

## Haemophilia A trial results 'mind-blowing'

By James Gallagher  
Health and science correspondent, BBC News

🕒 14 December 2017

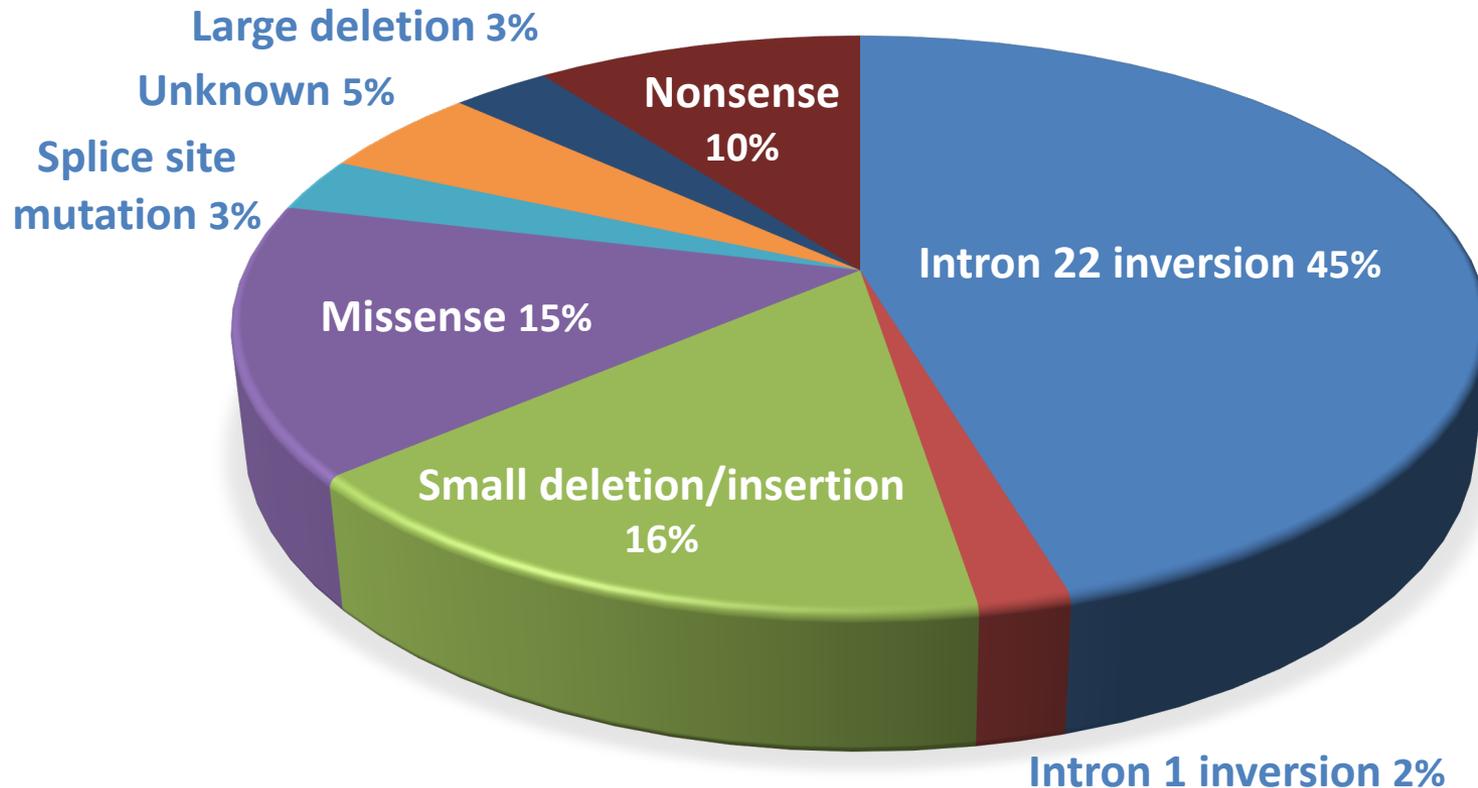
[f](#) [🐦](#) [💬](#) [✉](#) [Share](#)



Father-of-two Jake Ormer was born with haemophilia A

*BBC News,  
December 2017*

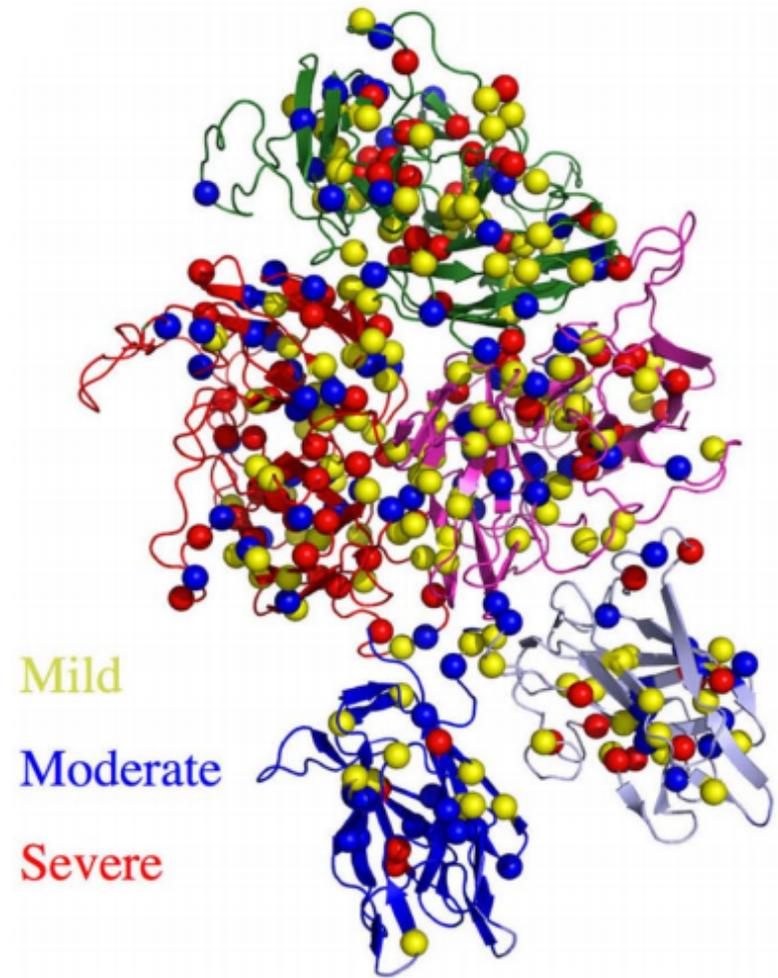
# F8 genotypes associated with HA



*Adapted from Gouw et al., Blood, 2012*

# FVIII missense mutations

- Total **977 missense mutations** (March 2018)
- Associated with **mild, moderate and severe haemophilia**
- ~10-15% develop **inhibitors** (anti-replacement FVIII antibodies)
- Accurate **risk stratification** has potential benefits (e.g. can inform choices about alternative therapeutic options)



# At the self/non-self boundary

A **single amino-acid substitution** difference will be detected by the immune system provided it occurs within at least one **spanning peptide** that:

- a) **Binds** to an MHC molecule long enough to be seen
- b) Forms a **novel peptide-MHC surface**
- c) There are T cells with **TCRs capable of binding to that surface**.

Note, however, that **detection is not sufficient** for the generation of inhibitors.

# Example missense mutation: R593C

tFVIII sequence spanning 593: ...FDEBRSWYLTENIQ**R**FLPNPAGVQLEDPE...

15-mer	MHC II binding core	predicted binding affinity, IC <sub>50</sub> (nmol/l)
FDEBRSWYLTENIQR	FDEBRSWYL	601.9
DEBRSWYLTENIQR <b>F</b>	YLTENIQR <b>F</b>	118.9
EBRSWYLTENIQR <b>FL</b>	YLTENIQR <b>F</b>	33.3
BRSWYLTENIQR <b>FLP</b>	<b>YL</b> TENIQR <b>F</b>	<b>39.4</b>
RSWYLTENIQR <b>FLPN</b>	YLTENIQR <b>F</b>	28.4
SWYLTENIQR <b>FLPNP</b>	YLTENIQR <b>F</b>	75.8
WYLTENIQR <b>FLPNPA</b>	IQR <b>FLPNPA</b>	103.9
YLTENIQR <b>FLPNPAG</b>	IQR <b>FLPNPA</b>	54.8
LTENIQR <b>FLPNPAGV</b>	IQR <b>FLPNPA</b>	21.3
TENIQR <b>FLPNPAGVQ</b>	<b>IQR</b> FLPNPA	<b>11.9</b>
ENIQR <b>FLPNPAGVQ</b> L	FLPNPAGVQ	9.0
NIQR <b>FLPNPAGVQ</b> LE	FLPNPAGVQ	8.9
IQR <b>FLPNPAGVQ</b> LED	FLPNPAGVQ	9.6
QR <b>FLPNPAGVQ</b> LED <b>P</b>	FLPNPAGVQ	16.7
R <b>FLPNPAGVQ</b> LED <b>PE</b>	FLPNPAGVQ	33.8

multiple predicted binding peptides spanning R593 with HLA-DRB1\*0101

Source: Hart et al., Haematologica, 2019

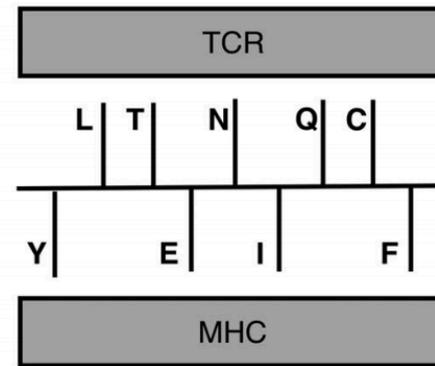
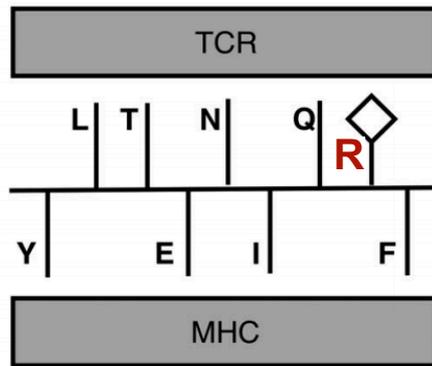
# pMHC surface novelty with R593C

MHC II binding core

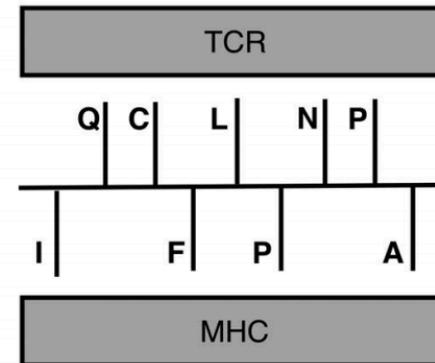
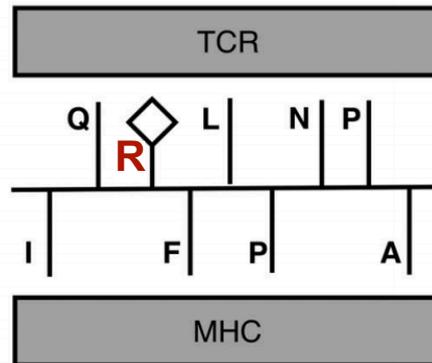
tFVIII peptide

endogenous FVIII peptide

**YLTENIQRF**

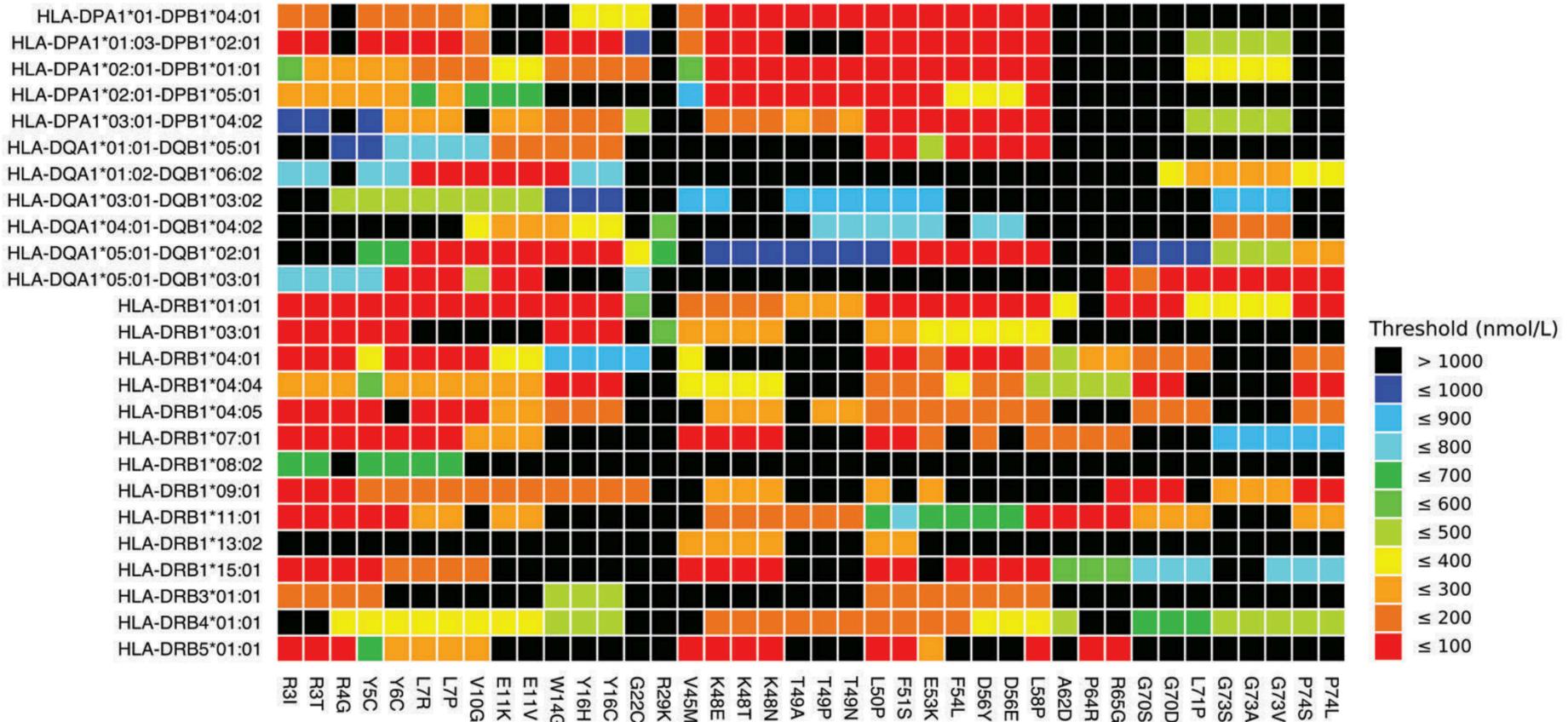


**IQRFLPNPA**



Source: Hart et al., Haematologica, 2019

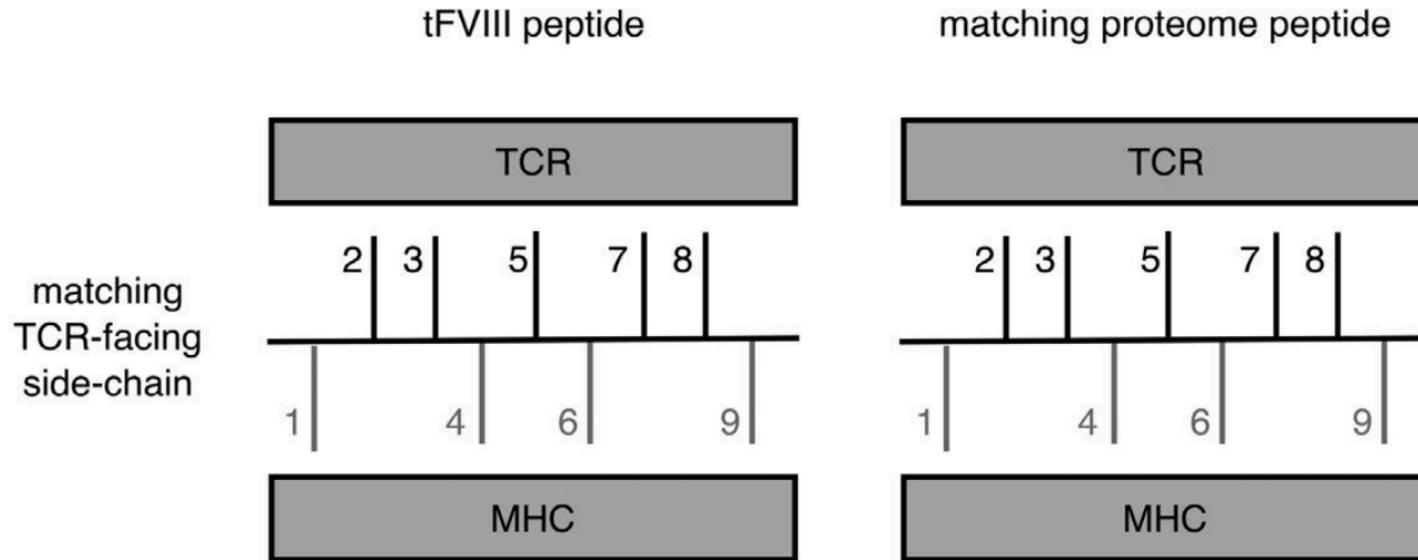
# Heatmap of inhibitor risk (I)



Percentage of **risk-associated combinations** = 49%

Source: Hart et al., Haematologica, 2019

# Proteome scanning



**Complete human proteome** from Ensembl contains over 100,000 proteins (including alternative isoforms) and **11,272,502 unique 9-mers**

*Source: Hart et al., Haematologica, 2019*

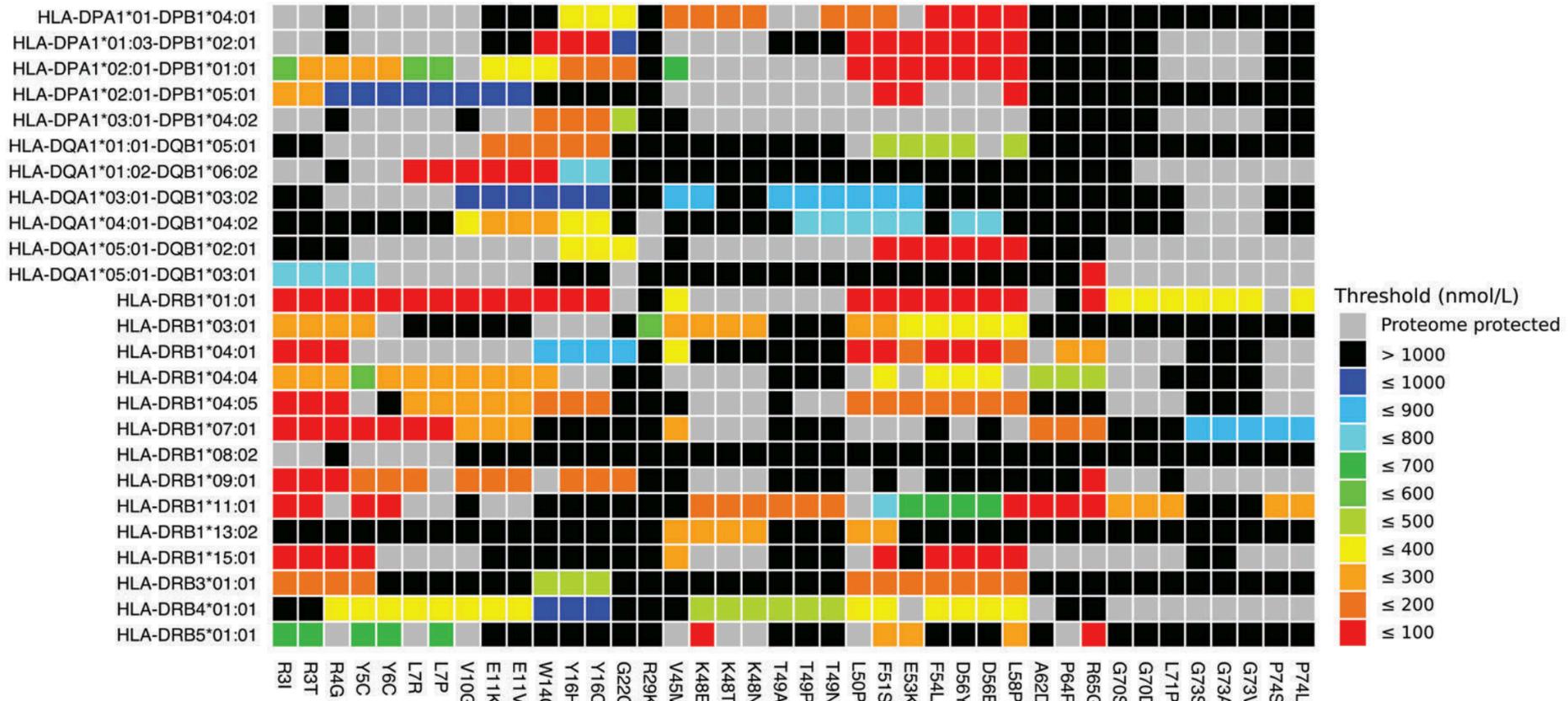
# Proteome scanning with R593C

Peptide **IQRFLNPA** has the following TCR-facing pattern:  
**XQRXLNPX**

15-mer	MHC II binding core	predicted binding affinity, IC <sub>50</sub> (nmol/l)
<b>tubulin polyglutamylase matches to pattern <b>XQRXLNPX</b> :</b>		
SGRAASFQRELNNPL	FQRELNNPL	26.3
GRAASFQRELNNPLK	FQRELNNPL	13.8
RAASFQRELNNPLKR	FQRELNNPL	7.9
AASFQRELNNPLKRM	FQRELNNPL	5.3
ASFQRELNNPLKRMK	FQRELNNPL	6.4
SFQRELNNPLKRMKE	FQRELNNPL	8.2
FQRELNNPLKRMKEE	FQRELNNPL	11.3

Source: Hart et al., *Haematologica*, 2019

# Heatmap with proteome scanning

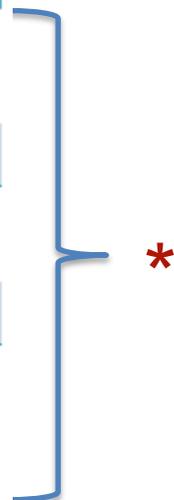


Percentage of **risk-associated combinations** = 31%

Source: Hart et al., Haematologica, 2019

# Proteins with most cross-matches

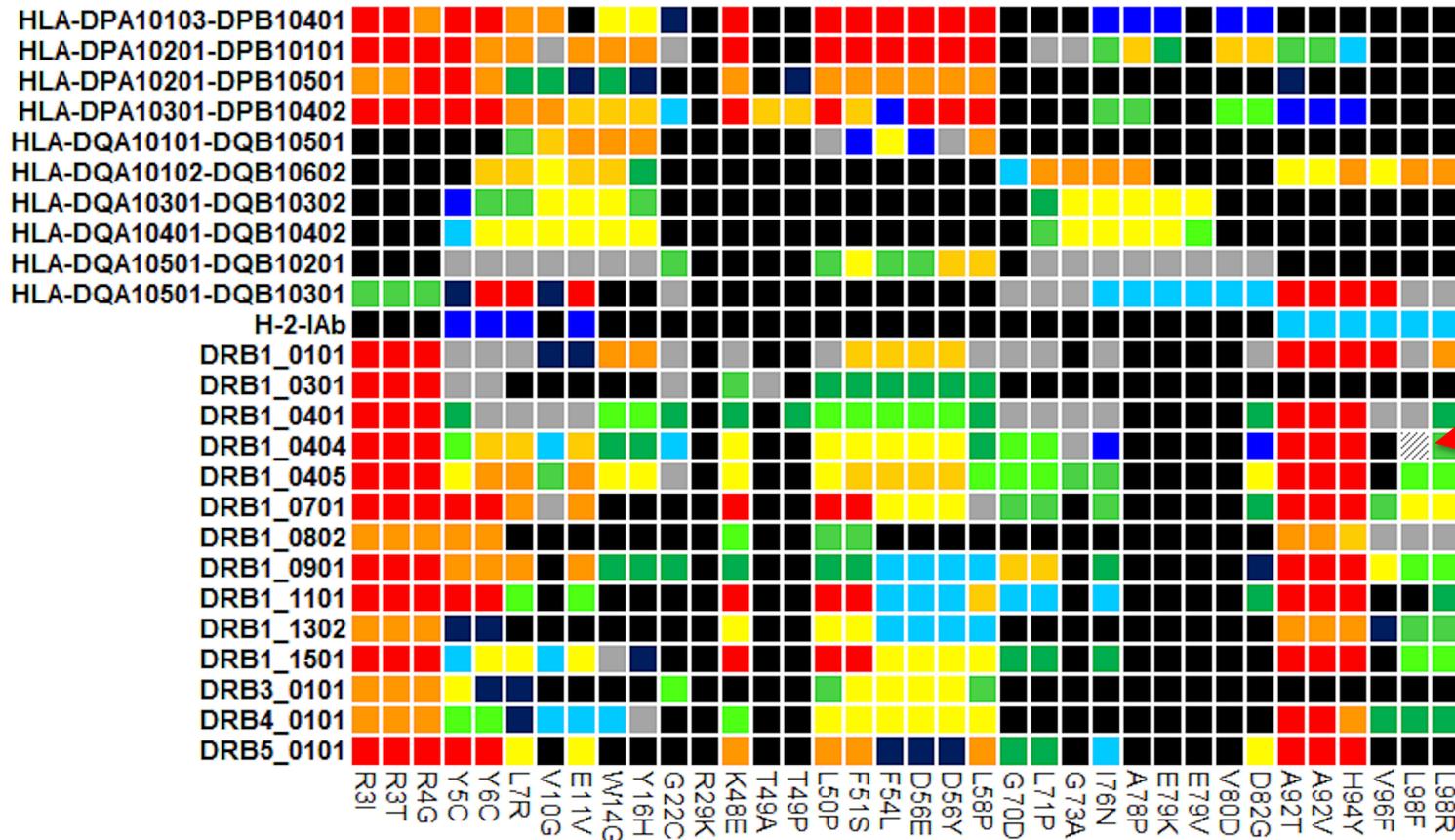
UniProt ID	Protein name	Protected peptide count
P12259	Coagulation factor V	640
Q6MZM0	Hephaestin-like protein 1	457
P00450	Ceruloplasmin	437
Q9BQS7	Hephaestin	389
P00451	Coagulation factor VIII [match to different, but homologous, location within the protein]	251
O75445	Usherin	150
Q14585	Zinc finger protein 345	142
Q14587	Zinc finger protein 268	134



\* all proteins with **copper-binding sites**

Source: Hart et al.,  
*Haematologica*, 2019

# Taking SNPs into account



SNPs with  $>5\%$  population frequency **reduced predicted risk by only 0.1%**

*MRes student Jacob Househam*

# Computational Challenges

# Antigen presentation and binding

**What can we predict?** We have always used the “Denmark tools” from **DTU** (now part of the Department of Health Technology):

## Class I:

NetMHCpan 4.0: predicts binding strength

NetCTLpan 1.1: predicts cleavage, TAP transport + binding strength

## Class II:

NetMHCIIpan 3.2: predicts binding strength + binding register



# Antigen binding thresholds (class I)

There is controversy about **absolute vs. relative** (percentile rank) thresholds. This is reflected in the recommendations of the **IEDB** website. **MHC class I:**

- 1) “Currently recommends” making selections based on **percentile rank of  $\leq 1\%$**  for each MHC allele/length combination (wrt to reference set of 1 million random natural sequences)
- 2) “Alternatively” use  $IC_{50}$  binding threshold of **500 nM**
- 3) Recently, “a paper from our group showed that... **MHC-specific thresholds should be used.**”

# Antigen binding thresholds (class 2)

These are the **IEDB recommendations** for **MHC class II**:

- 1) **percentile rank** of top 10%
- 2) **IC<sub>50</sub> threshold** of 1,000 nM

Note the following advice for **both class I and class II** under **Interpreting predicted results**:

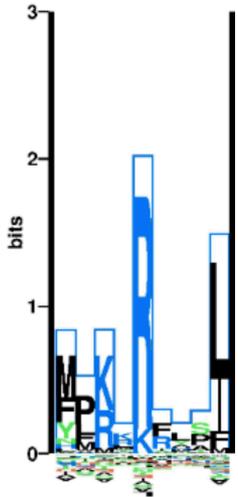
“As a rough guideline, peptides with IC<sub>50</sub> values <50 nM are considered **high affinity**, <500 nM **intermediate affinity** and <5000 nM **low affinity**.”



# Peptide anchors

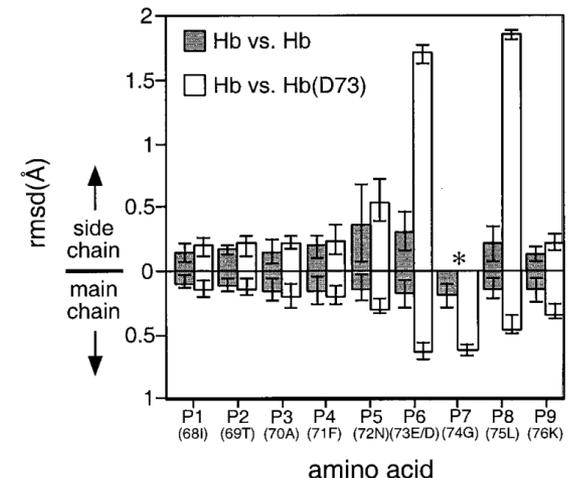
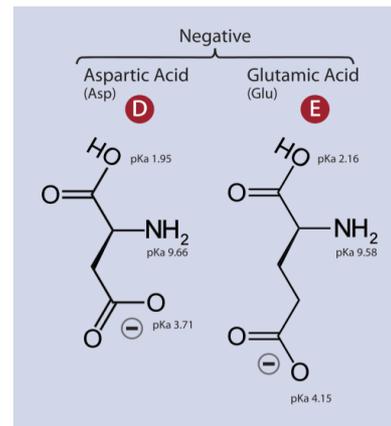
Which are the **anchoring positions**?

“Canonical” MHC class I & II molecules have pockets at positions 2 + 9 and 1 + 4 + 6 + 9 respectively. But some have additional/ alternative pockets.



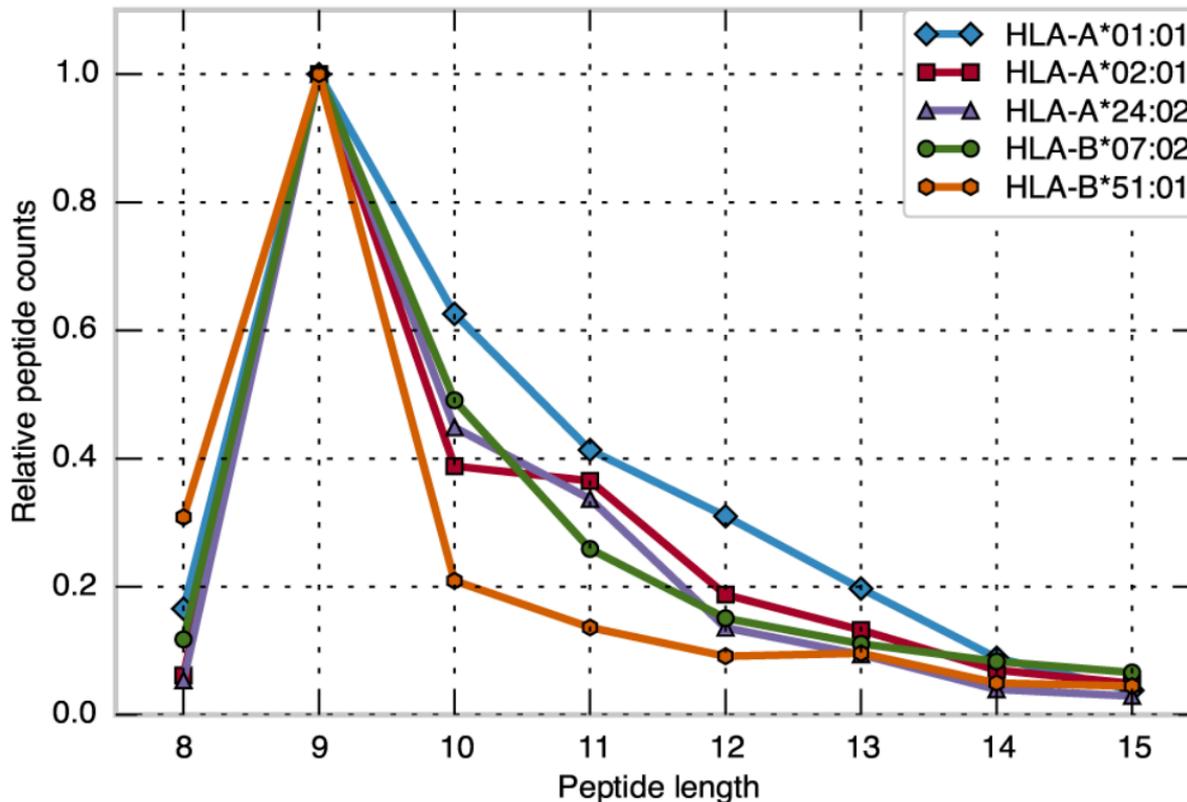
HLA-B\*08:01 using NetMHCpan v2 (Source: MHC Motif Viewer)

Evidence that mutations to anchoring residues may introduce novelty via **main chain displacement** (Kersh et al., Jol, 2001)



# Challenges: class I peptide lengths

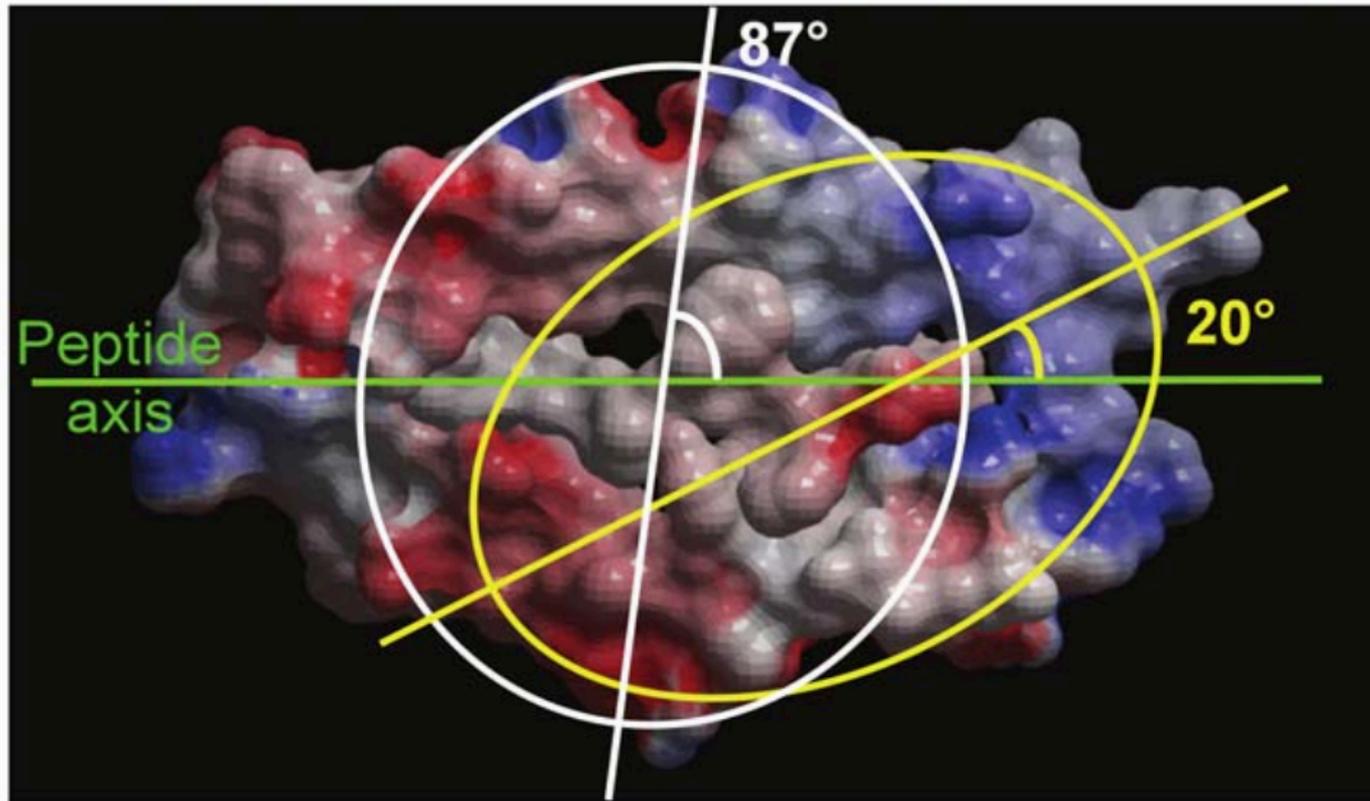
Most people (including members of my group) focus on **9-mers**.



Length of naturally-presented peptides for 5 HLAs

*Trolle et al., Jol, 2016*

# Challenges: TCR binding orientation



Source: Khan & Ranganathan, PLoS ONE, 2011

# Acknowledgements



## My Group

Naz Uzun

Stuart Skelton

Jacob Househam

Alison Kakoschke

Zainab Aziz

William Lees

## Collaborators

### Haemophilia A

Dr Dan Hart (QMUL, Barts)

### HCC

Dr Shilpa Chokshi (Inst  
Hepatology)



The AIRR Community

