

# EpiVax **Novel immunogen design elicits increased protection against avian H7N9 influenza that is associated with mobilization of seasonal influenza T cell memory**

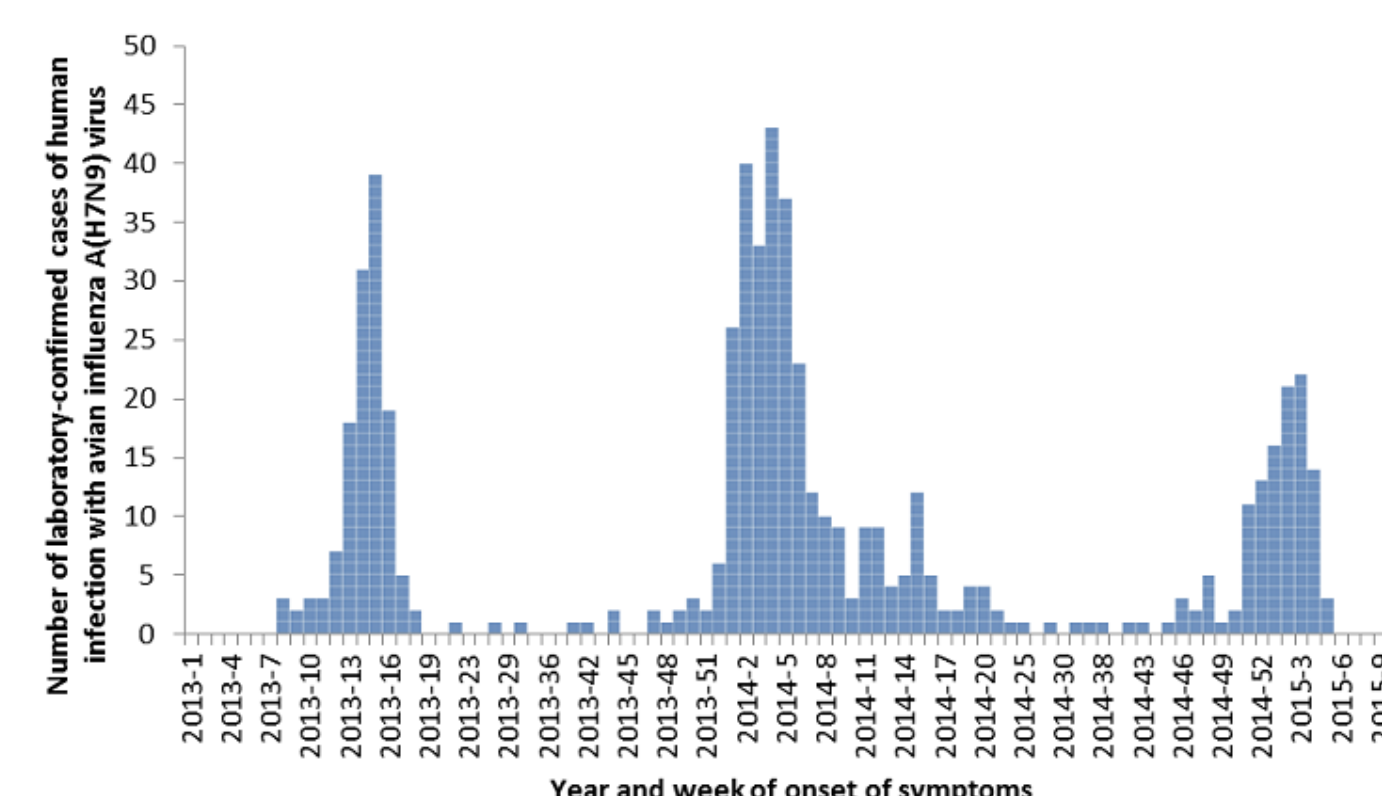
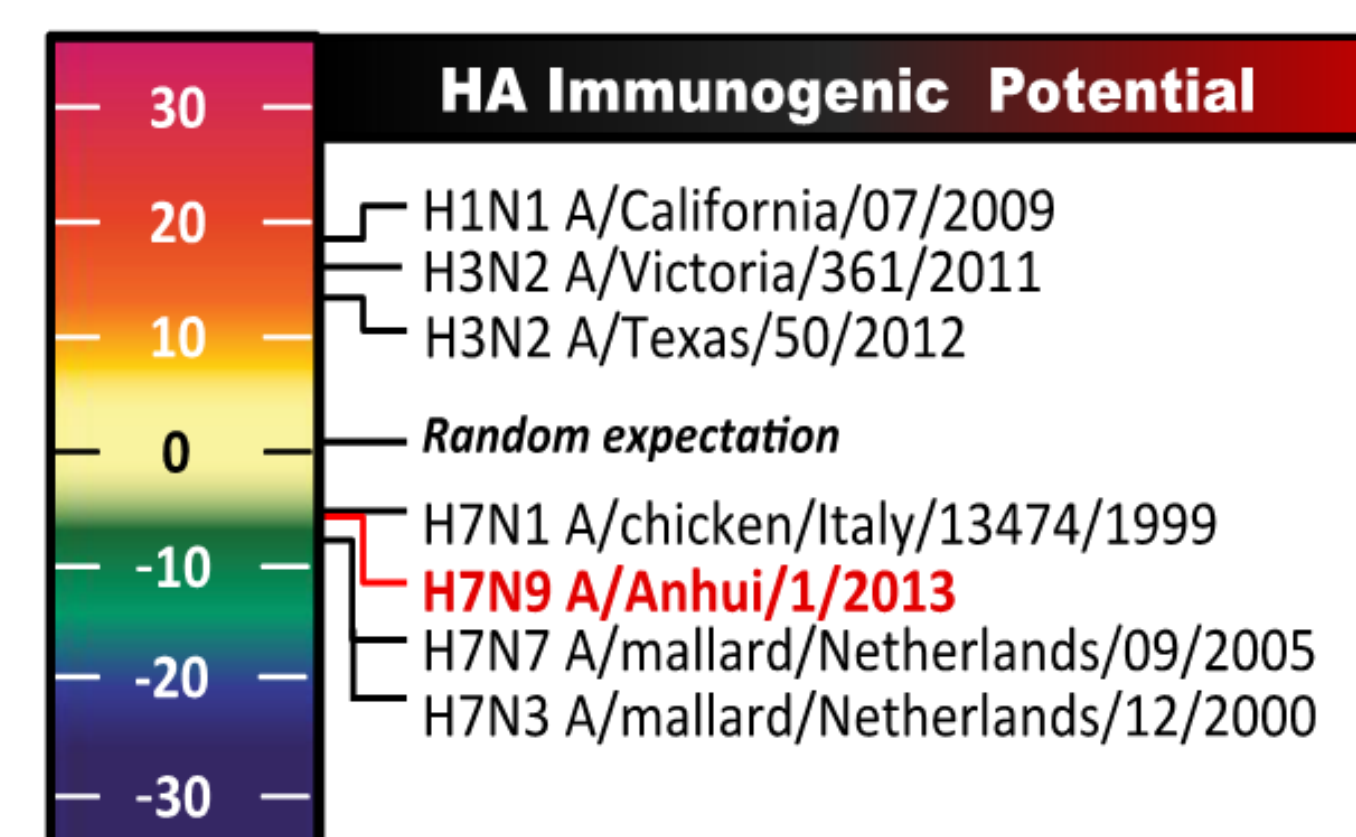
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## Introduction

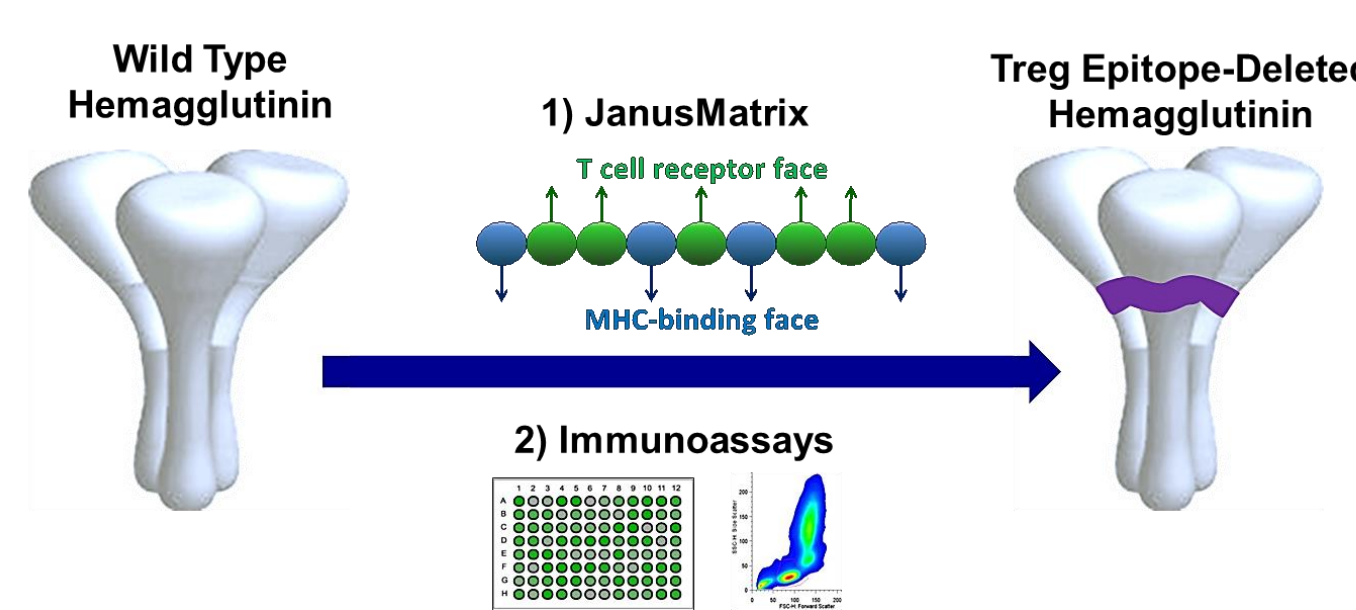
A new avian-origin influenza virus emerged near Shanghai in February 2013. Human-to-human transmission of avian-origin H7N9 influenza A has been limited to a few family clusters, but the high mortality rate (27%) associated with human infection has raised concern about the potential for this virus to become a significant human pathogen.

- Previously, H7 HA-containing vaccines have been poorly immunogenic.
- We used well-established immunoinformatics tools to estimate the immunogenic potential of H7N9 proteins.
- HA proteins derived from human-derived H7N9 strains contain fewer T cell epitopes than most other circulating strains of influenza.
- Conservation of T cell epitopes with other strains of influenza was very limited.
- Based on our analysis, avian-origin H7N9 2013 appears to be a "stealth" virus.
- Improved vaccines for emerging influenza are urgently needed.



## Methods

- Sequence analysis of the two faces of H7N9 T cell epitopes revealed sequences with potential to activate Tregs.
- Immunoassays confirmed an H7N9 Hemagglutinin (HA) sequence that increases the frequency of Tregs and suppresses effector responses (Liu *et al.* (2015) *Human Vaccines & Immunotherapeutics*, 11:2241-2252).
- Site-specific modifications were engineered in H7N9 HA to delete this Treg-activating epitope and to harness memory CD4 effector T cells.**
- OPT1 (three amino acid substitutions replaced a reported regulatory T cell (Treg) epitope with a highly conserved and broadly reactive CD4+ T cell epitope from H3-HA) and OPT2 (integrates six H3-HA CD4+ T cell epitopes, one Treg epitope removed).

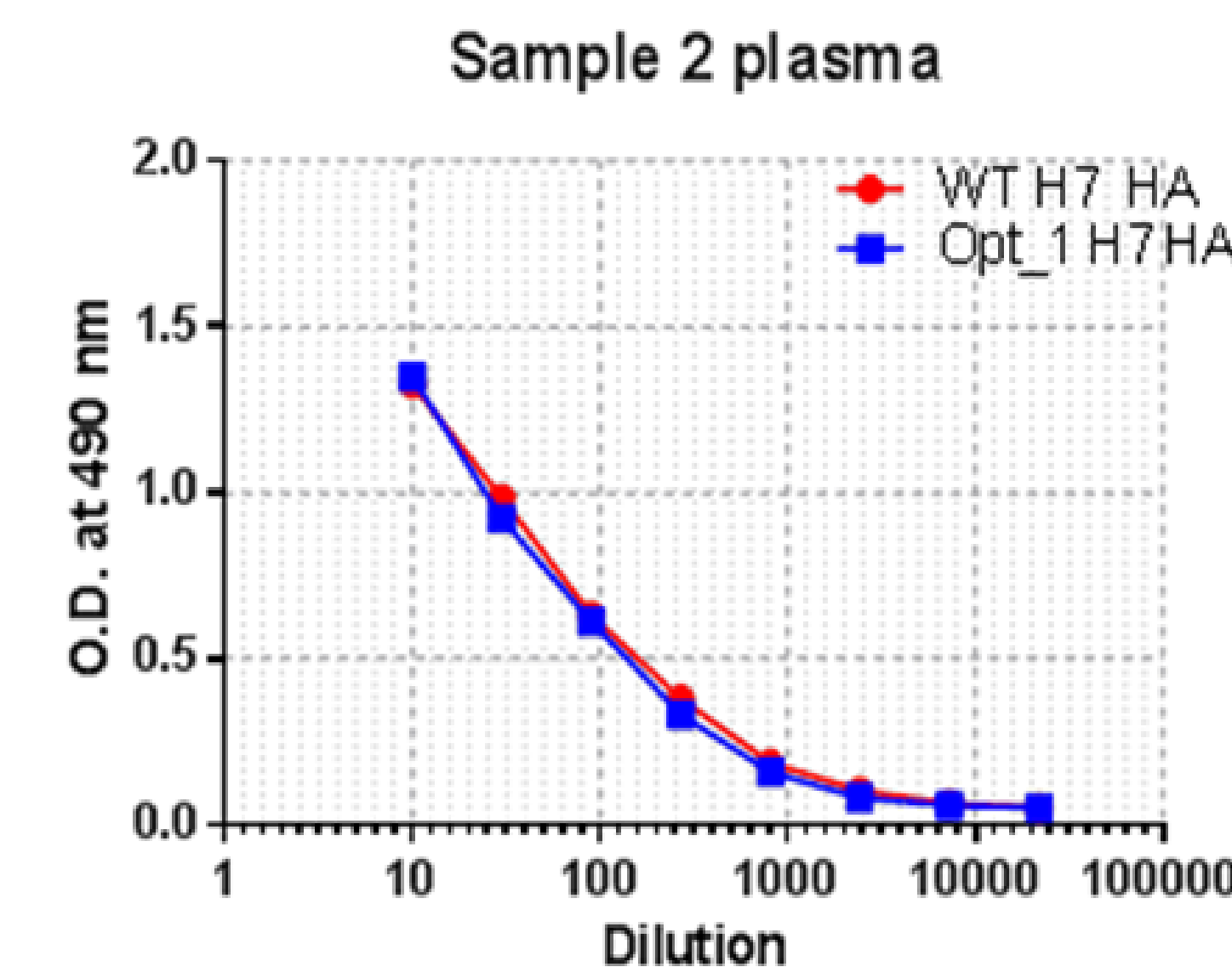
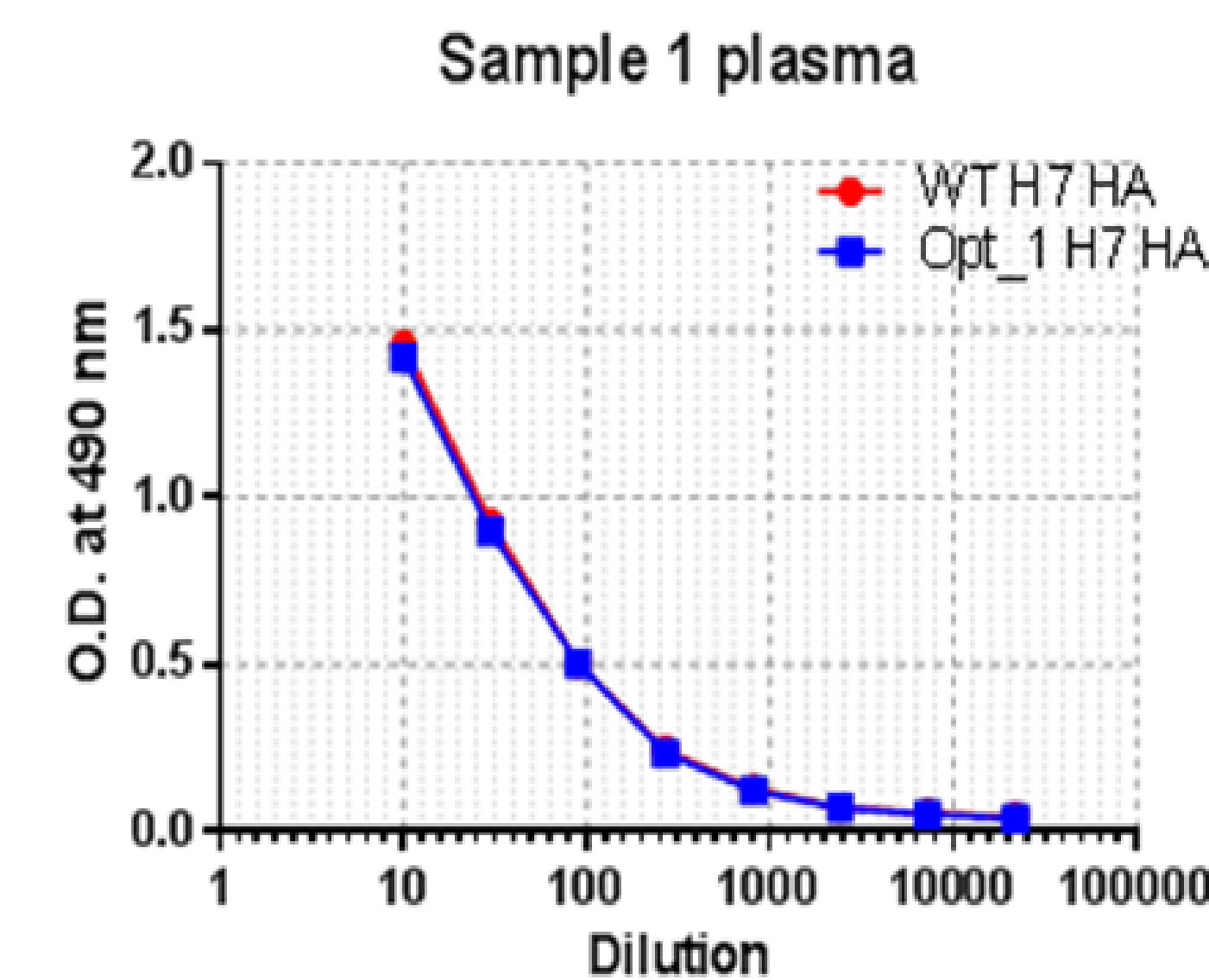
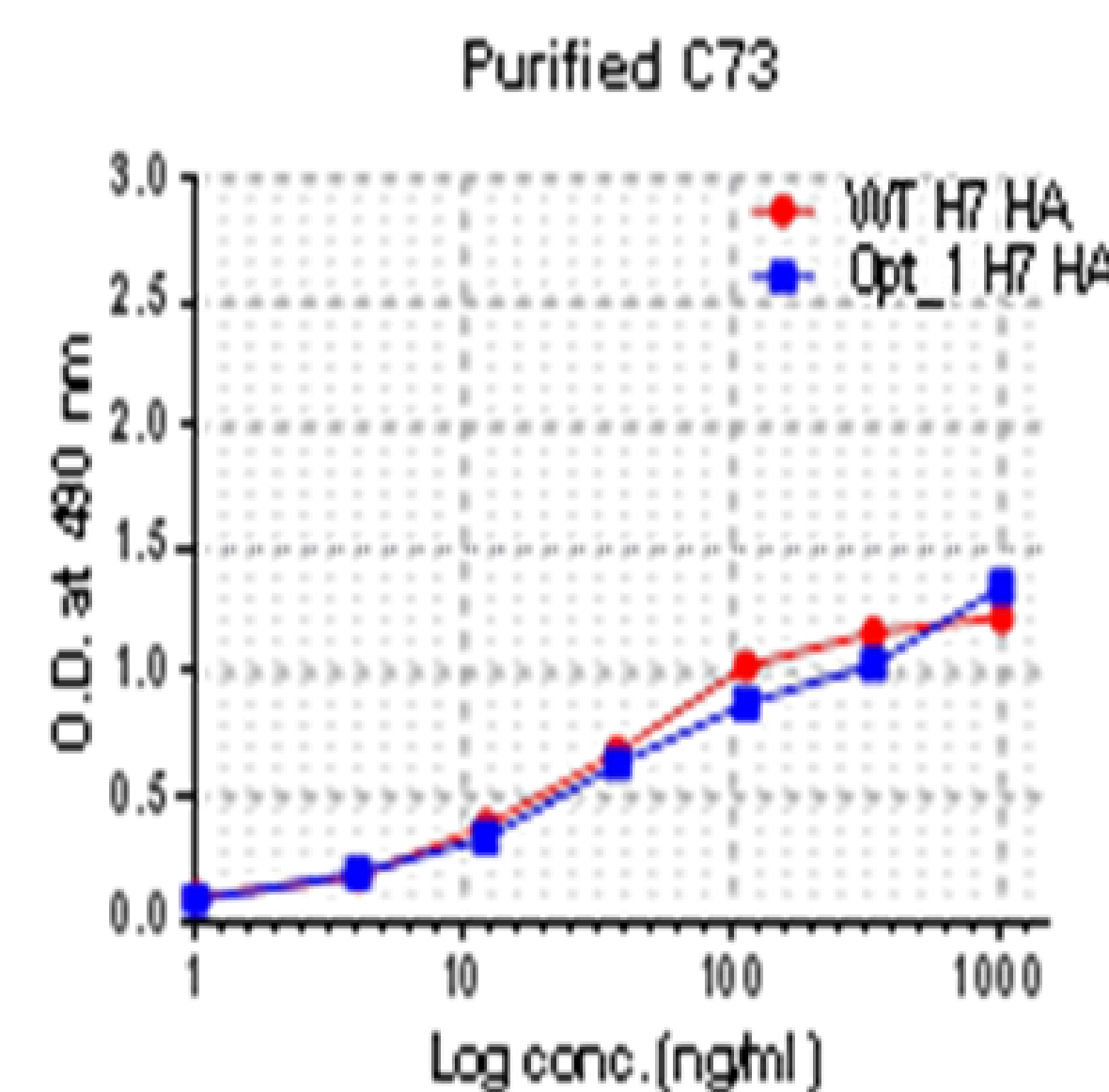
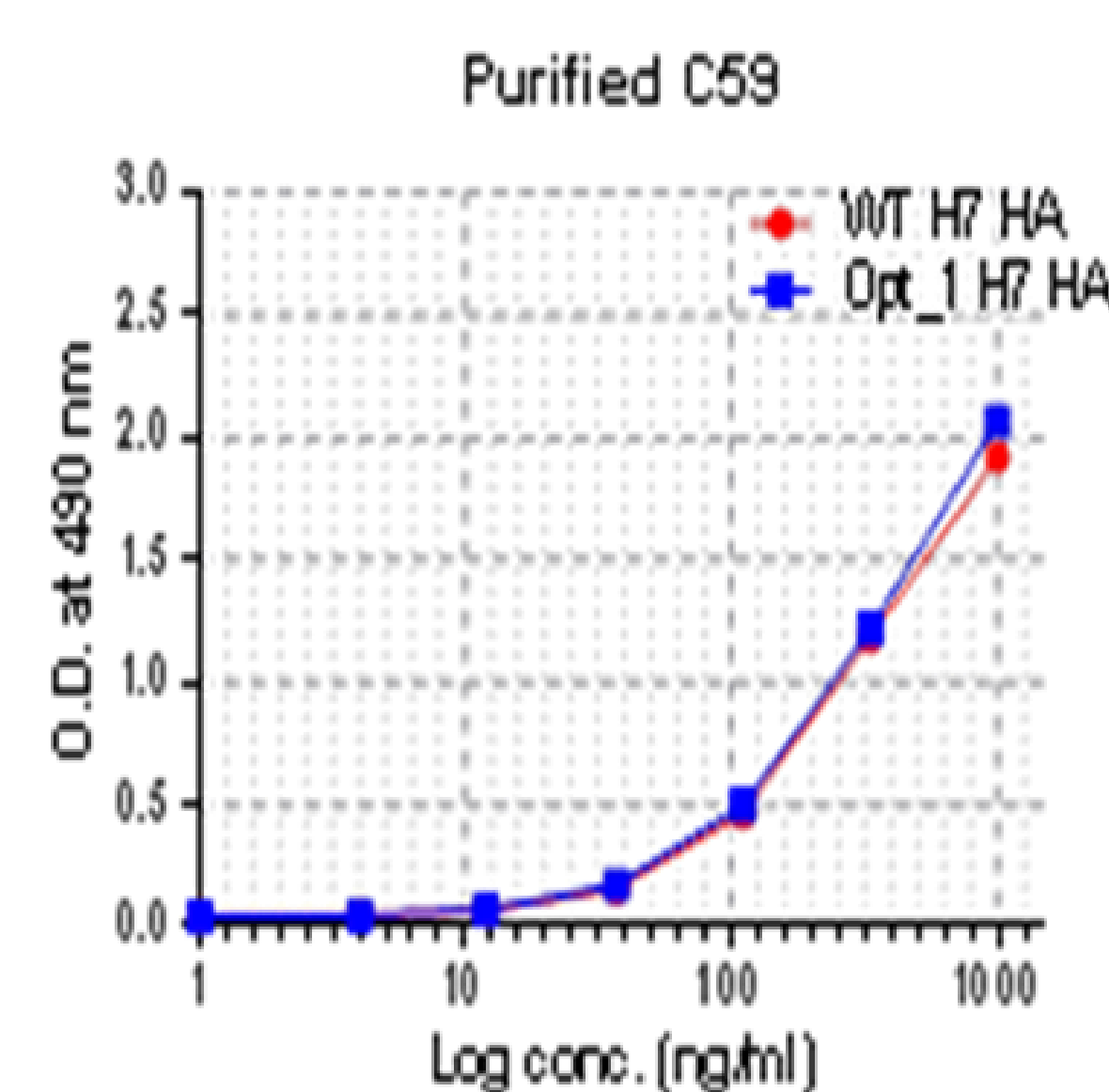


## Immunization

Vaccination studies were carried out in HLA-DR3 transgenic mice pre-immune to H3N2 (A/Hong Kong/4108/2014).

- Mice received two H3N2 exposures (mucosal and systemic), followed by three rHA vaccinations.
- Mice were primed and boosted twice IM with wildtype or engineered H7N9 rHA without adjuvant eight weeks post-H3N2 exposure.
- Mice were challenged IN with H7N9 virus (A/Anhui/1/2013) four weeks following boost and followed for weight loss and survival.

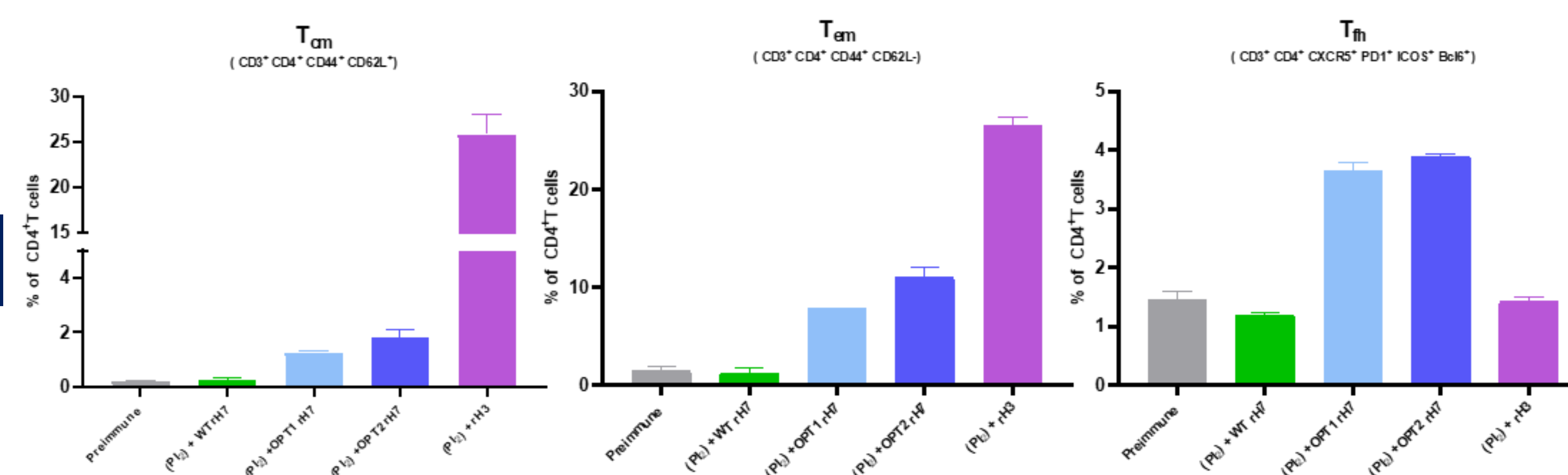
## Engineered H7-HA maintained antigenic structures



Previously identified monoclonal antibodies against WT rH7-HA also recognize Opt\_1 rH7-HA

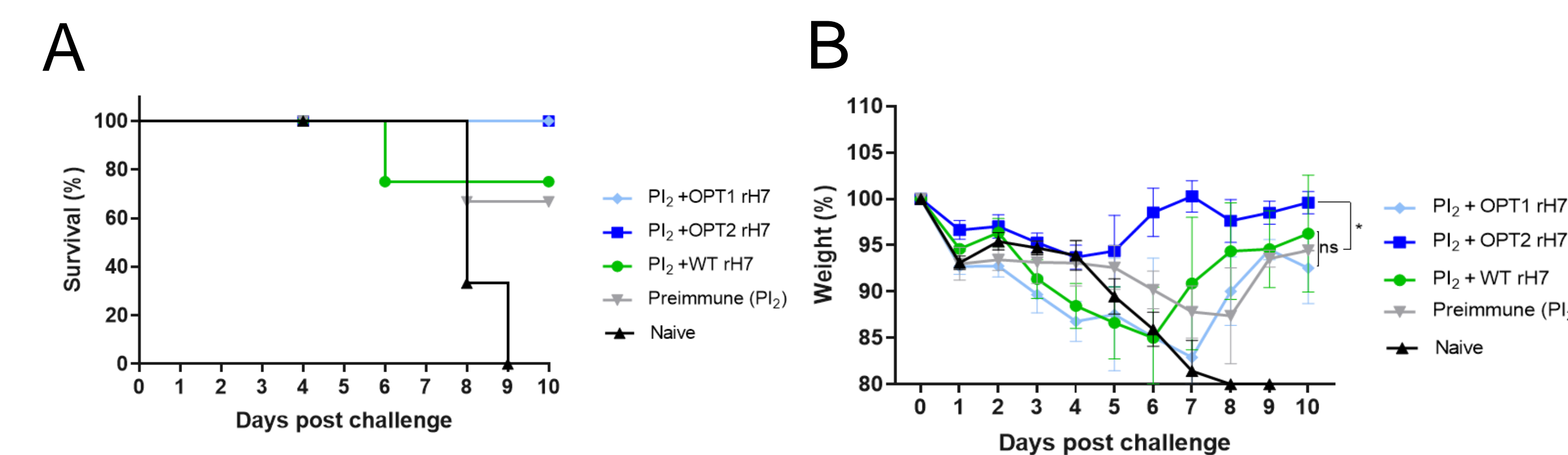
Antibodies from two patients with previously identified anti H7-HA antibodies also recognize Opt\_1 rH7-HA

## OPT1 and OPT2 rH7 enhance memory T cell response



Analysis by flow cytometry uncovered that, compared to preimmune controls, OPT1 and OPT2 vaccinations upregulated the population of Tcm (left), Tem (middle) and Tfh (right).

## OPT1 and OPT2 vaccines protect against lethal H7N9



OPTimized vaccines protected against lethal H7N9 virus infection while in wildtype vaccinated animals had a survival rate similar to unvaccinated pre-immune controls. OPT1 and OPT2 lowered average weight loss post-infection with OPT2 animals maintaining >95% of their weight.

## Conclusions

- Treg epitope deletion preserves H7N9 HA antigenicity.
- Epitope-driven approaches to vaccine design that involve careful consideration of the T cell subsets primed during immunization are a promising means of enhancing vaccine efficacy.
- Collectively, we find increased vaccine-induced protection against a pandemic influenza strain is associated with enhanced T cell immunity elicited by a novel immunogen design strategy that harnesses seasonal influenza CD4+ T cell memory.
- Structure-guided T cell epitope modification may also be useful to develop more effective vaccines against seasonal influenza strains.

## References

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