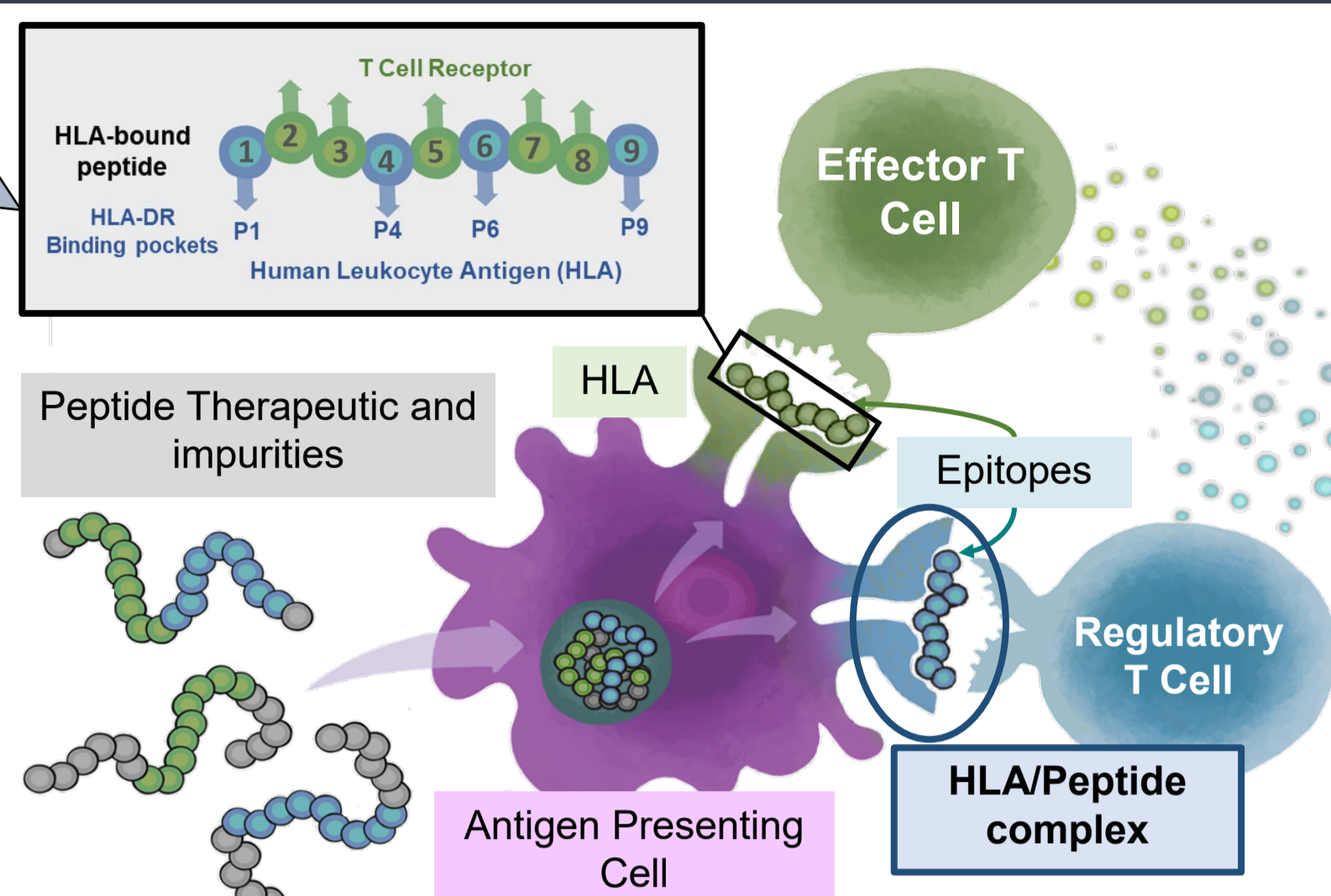


BACKGROUND INFORMATION

In silico prediction of T cell epitopes within a peptide drug candidate serves as an important 1st step for assessing immunogenicity.

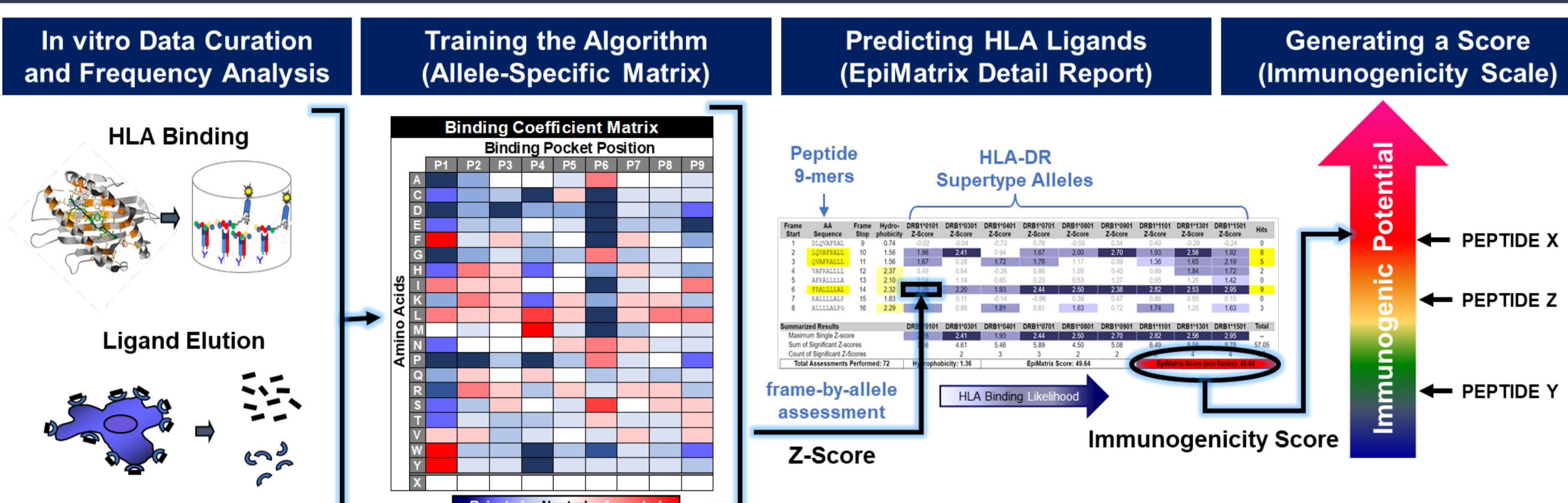
T cell epitopes bind HLA by a well-characterized interaction of amino acid side chains and pockets in the HLA-DR molecule binding groove.



Immunoinformatics tools, such as EpiMatrix, have been developed to screen natural amino acid sequences for peptides that will bind HLA.

In silico assessment of immunogenic potential allows for risk-based selection of best candidate peptides in further confirmatory in vitro, ex vivo and in vivo assays, thereby reducing the overall cost of immunogenicity evaluation.

Immunogenicity Scores are calculated from in silico algorithms trained on curated in vitro data for natural AA peptide sequences

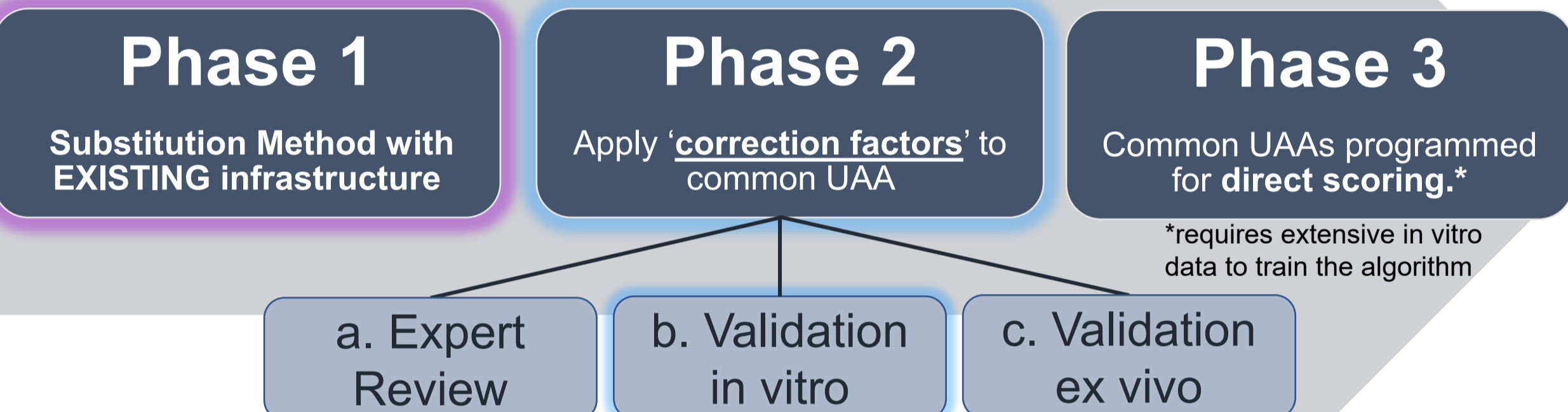


HLA binding properties of peptides containing unnatural amino acids (UAA) are not accurately estimated by most algorithms, to date.

UAA are often incorporated into peptide therapeutics to improve drug properties and commonly occur in synthetic peptide-related impurities. Both scenarios warrant the need for enhanced predictive algorithms.

OVERVIEW & ROAD MAP

Process to expand existing in silico immunogenicity prediction tools to handle sequences containing unnatural amino acids



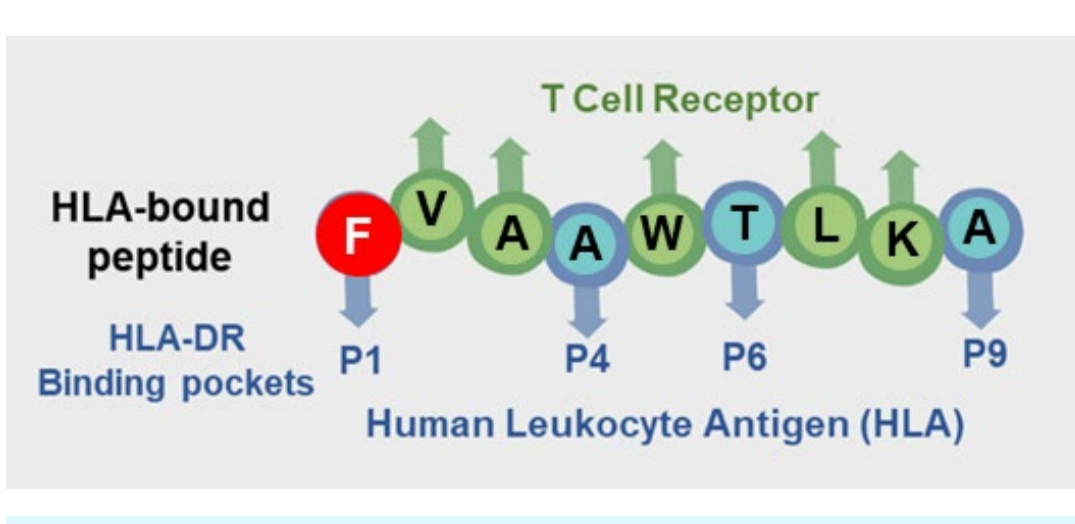
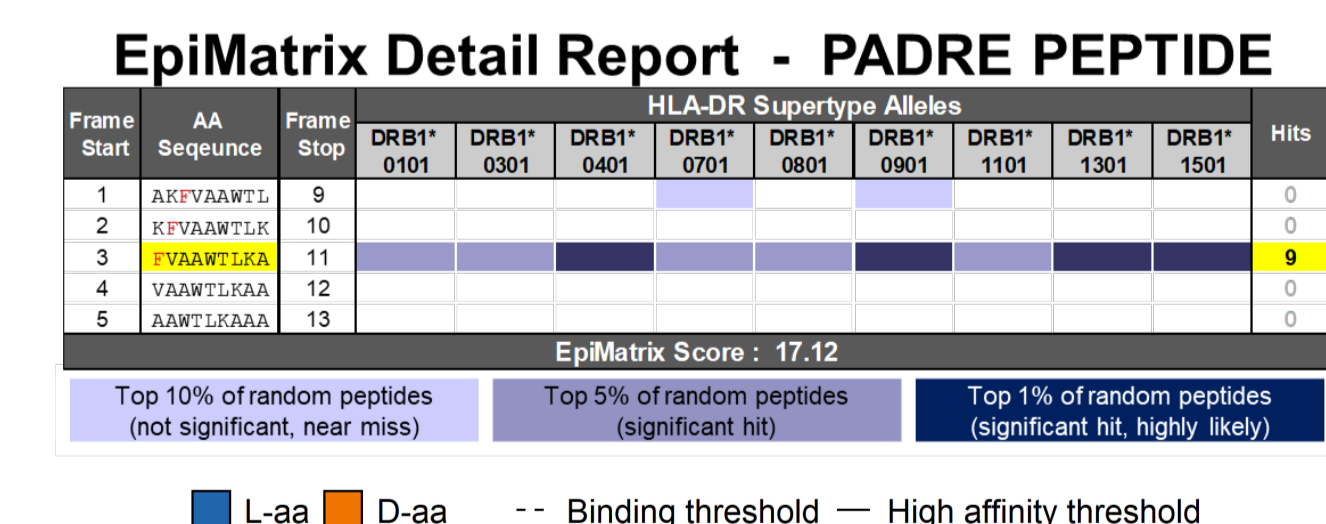
- Natural amino acid or placeholder substitutions must be applied to score peptides that include UAA with existing epitope mapping tools. (Phase 1)
- 'Correction factors' can be applied to the scores of natural amino acid substitutions to more accurately predict HLA binding for UAA-containing sequences. (Phase 2)
 - D-amino acids and side chain modifications that introduce 'bulk' are expected to negatively impact the HLA binding likelihood and can be modeled by introducing a deduction to the score of the closest matching natural L-amino acid. (Phase 2a)
 - These 'correction factors' will be further refined based on in vitro validation data. (Phase 2b,c)

PHASE 2: Apply "Correction Factors" to common UAA

- Expert Review**
Review of UAA side chain structure compared to closest matching natural AA and apply score deductions. (i.e. minimal, moderate, or significant).
- Validation in vitro**
HLA binding assays to compare known ligand sequences modified with selected UAA in HLA binding positions. See Below for example with D-AA.
- Validation ex vivo**
IVIP T cell assays to assess the impact of selected UAA on T cell recognition and immunogenic potentials.

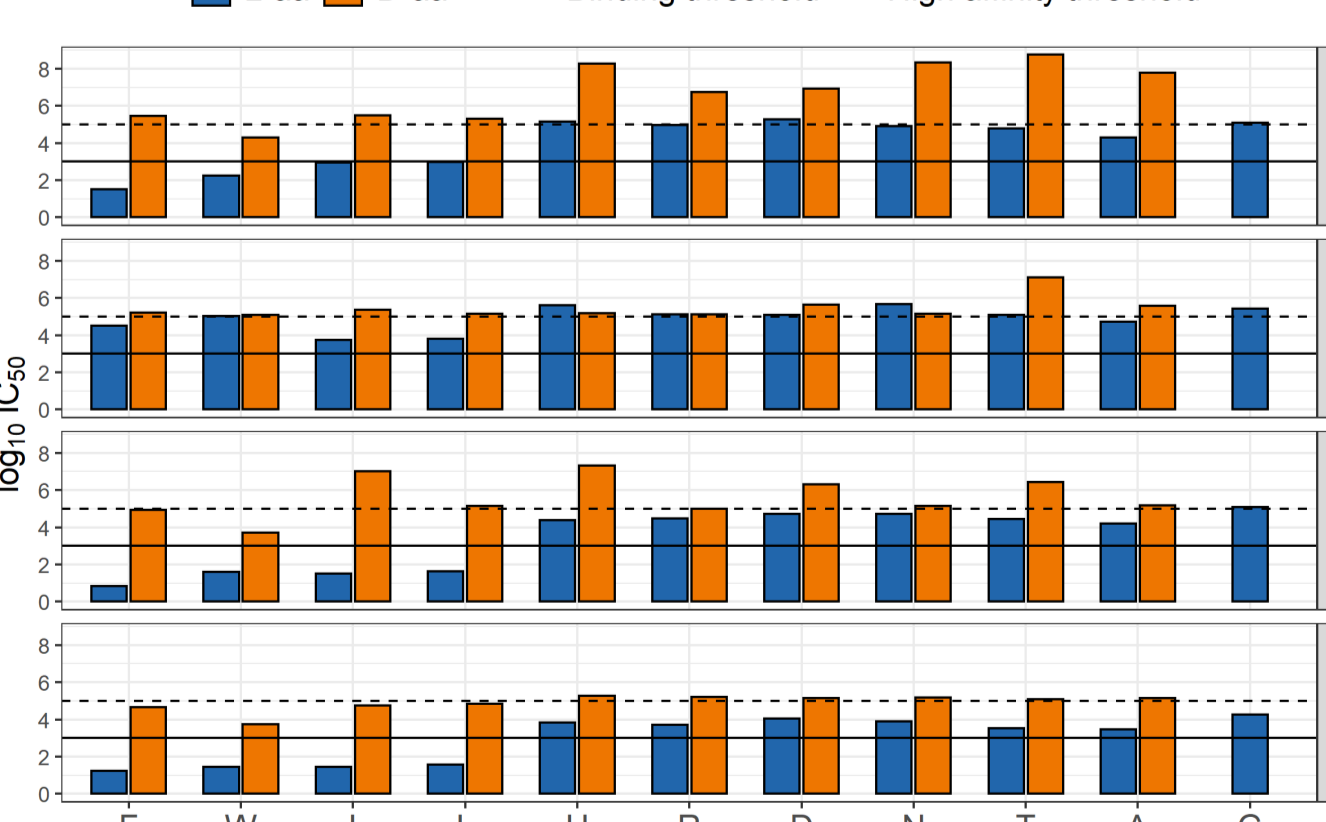
Preliminary in vitro data for D-amino acid 'correction factors'

HLA binding studies based on known ligands (such as PADRE) which have been modified to contain UAAs in HLA anchoring positions (1,4,6,9) can be used to estimate the binding potentials of commonly encountered UAA and "correct" in silico estimates of binding based on their naturally-occurring counterparts.



D-amino acids disrupt HLA binding compared to their corresponding L-amino acid isomer when substituted in HLA binding position 1 of a known promiscuous HLA-DR binding peptide (PADRE).

Similar studies are underway assessing the impact of D-AA in other positions. These studies will enable calculation of correction factors for adjusting in silico HLA binding estimates for D-AA, and to further adjust the binding predictions for D-AA by allele and amino acid group.



PHASE 1 : Substitution Method with Existing in silico Algorithms

Three Steps to Select a Best Proxy Substitution for the Unnatural Amino Acid

- Neutral Placeholder**
Replace the UAA with a neutral placeholder X. In EpiMatrix, amino acid "X" has a coefficient of 0 and is assumed to neither promote nor detract from binding.
- Replacement Analysis**
Replace the UAA with each of the 20 natural L-amino acids to establish the extent to which variation at this position can have an impact on the binding potential of the input peptide and identify which, if any, substitutes are likely to promote or significantly detract from HLA binding potential.
- Structural Proxy**
Review the structural/chemical properties of the UAA side chain and, if applicable, replace with the closest matching natural L-amino acid.

Example : Semaglutide API and D-Amino Acid Impurities

Selecting suitable replacements for the UAA in Semaglutide API

Aminoisobutyric Acid (Aib)

- Neutral placeholder:** Replace UAA (Aib) with X
- Replacement Analysis:** Replace X with all 20 natural L-amino acids
- Structural Proxy:** Choose best-matching natural L-amino acid based on structural and/or chemical properties

Aminoisobutyric acid (Aib)

- Narrow range of scores → little impact on HLA binding potential
- Replace with structural proxy, Ala.

EpiMatrix analysis for Semaglutide API

K(OEG-OEG-γGlu-C18diacid)

- Neutral placeholder:** Replace UAA (K(OEG-OEG-γGlu-C18 diacid)) with X
- Replacement Analysis:** Replace X with all 20 natural L-amino acids
- Structural Proxy:** Choose best-matching natural L-amino acid based on structural and/or chemical properties

K(OEG-OEG-γGlu-C18diacid)

- Narrow range of scores → little impact on HLA binding potential
- Expected to disrupt potential HLA binding, replace with low affinity placeholder, Z.

EpiMatrix Detail Report - SEMAGLUTIDE

Frame Start	AA Sequence	Frame Stop	DRB1*0101	DRB1*0301	DRB1*0401	DRB1*0701	DRB1*0801	DRB1*0901	DRB1*1101	DRB1*1301	DRB1*1501	Hits
7	HAEGTFTSDVSSYLEGQAAEFIAWLVRGRG	15	0	0	0	0	0	0	0	0	0	0
8	AEFTTSPD	16	0	0	0	0	0	0	0	0	0	0
9	KEFTTSPD	17	0	0	0	0	0	0	0	0	0	0
10	QFTTSPD	18	0	0	0	0	0	0	0	0	0	0
11	TFTTSPD	19	0	0	0	0	0	0	0	0	0	0
12	YFTTSPD	20	0	0	0	0	0	0	0	0	0	0
13	SEFTTSPD	21	0	0	0	0	0	0	0	0	0	0
14	SNFTTSPD	22	0	0	0	0	0	0	0	0	0	0
15	DVSSYLEGQ	23	0	0	0	0	0	0	0	0	0	0
16	VSSYLEGQA	24	0	0	0	0	0	0	0	0	0	0
17	SYLSEGQA	25	0	0	0	0	0	0	0	0	0	0
18	SYLSEGQAI	26	0	0	0	0	0	0	0	0	0	0
19	YLEGQAEE	27	0	0	0	0	0	0	0	0	0	0
20	LEGQAEEF	28	0	0	0	0	0	0	0	0	0	0
21	EQAAEEFT	29	0	0	0	0	0	0	0	0	0	0
22	QAAEEFTI	30	0	0	0	0	0	0	0	0	0	0
23	QAEEFTIA	31	0	0	0	0	0	0	0	0	0	0
24	AAEEFTIWL	32	0	0	0	0	0	0	0	0	0	0
25	AEFTIWLV	33	0	0	0	0	0	0	0	0	0	0
26	FTIWLVR	34	0	0	0	0	0	0	0	0	0	0
27	FIWLVRGR	35	0	0	0	0	0	0	0	0	0	0
28	FWLVRGR	36	0	0	0	0	0	0	0	0	0	0
29	WLVRGRG	37	0	0	0	0	0	0	0	0	0	2

EpiMatrix Score: -0.25

Top 10% of random peptides (not significant, near miss) | Top 5% of random peptides (significant hit) | Top 1% of random peptides (significant hit, highly likely)

*note: semaglutide peptide numbering is relative to hGLP-1 (7-37)

Semaglutide D-amino acid impurity analysis examples

D-His in position 7*

- Neutral placeholder:** Replace UAA (D-His) with X
- Replacement Analysis:** Replace X with all 20 natural L-amino acids
- Structural Proxy:** Choose best-matching natural L-amino acid based on structural and/or chemical properties

D-His7

- No change in score
- modification will not impact HLA binding

D-Phe in position 12*

- Neutral placeholder:** Replace UAA (D-Phe) with X
- Replacement Analysis:** Replace X with all 20 natural L-amino acids
- Structural Proxy:** Choose best-matching natural L-amino acid based on structural and/or chemical properties

D-Phe12

- Wide range of scores, all lower than baseline score
- modification will disrupt HLA binding

In Vitro HLA Binding Study Confirms in Silico Predictions

Baseline API

HLA DRB1*0401

Moderate Affinity (IC50=4,230)

Impurity

HLA DRB1*0401

Moderate Affinity (IC50=4,120)

D-His7

HLA DRB1*0401

Moderate Affinity (IC50=8,750)

D-Phe12

HLA DRB1*0401

Non-Binder

TAKE HOME MESSAGE

- In silico risk assessment of peptides and their related impurities is an important first step to understanding the immunogenic potential of a given therapeutic, but in silico immunogenicity prediction algorithms are limited to natural amino acid sequences.
- A three-phased approach for the eventual incorporation of common unnatural amino acids into immunoinformatic toolkits includes: first, a substitution-based method enabling in silico immunogenicity risk assessment for sequences containing unnatural amino acids, and second, the use of in vitro HLA binding and ex vivo T cell assays in the development of 'correction factors' that can be applied to in silico 'scores' for common unnatural amino acids, for more accuracy in predictions.

FUNDING & REFERENCES

Some of the data presented was funded in part by FDA Contract # 75F40120C00157.
Mattei AE, Gutierrez AH, Martin WD, Terry FE, Roberts BJ, Rosenberg AS and De Groot AS (2022), Front. Drug. Discov. 2:952326. doi: 10.3389/fddv.2022.952326